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USPT.JPAB.EPAB.DWPL.TDBD	110 and 111 and 112 and 114 and 117	11	L18
USPT.JPAB.EPAB.DWPL.TDBD	ampicillin	11779	L17
USPT.JPAB.EPAB.DWPL.TDBD	110 and 111 and 112 and 114	16	L16
USPT.JPAB.EPAB.DWPL.TDBD	(phenylglycine amide) or (d-phenylglycine amide)	58	L15
USPT.JPAB.EPAB.DWPL.TDBD	(sulfuric acid) or h2so4	186645	L14
USPT.JPAB.EPAB.DWPL.TDBD	(sulfuric acid) or h2so4	30723	L13
USPT.JPAB.EPAB.DWPL.TDBD	(d-phenylglycine) or phenylglycine	3390	L12
USPT.JPAB.EPAB.DWPL.TDBD	enzymS5	176652	L11
USPT.JPAB.EPAB.DWPL.TDBD	(6-aminopenicillanic acid) or 6-APA	1962	L10
JPAB.EPAB.DWPI	CN-1165032-\$.did.	0	L9
JPAB.EPAB.DWPI	jp-05204120-\$.did.	2	L8
JPAB.EPAB.DWPI	jp-02240026-\$.did.	2	L7
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The novel Amphoteric ester or its acid addition salt is prepared by (1) reacting a corresponding carboxylic acid (VI) or its reactive derivative, (2) with a compound of the formula $\text{H}_2\text{N}^+\text{R}_1\text{N}^+\text{R}_2$ wherein R_1 and R_2 are as defined above, and R is a C_1 - C_6 alkyl, or reacting a compound of the formula $\text{H}_2\text{N}^+\text{R}_1\text{N}^+\text{R}_2$ wherein R_1 and R_2 are as defined above, or its acid addition salt with a corresponding carboxylic acid (VI) or its reactive derivative, (2) thereafter, if required, when the resulting compound has the protected amino group or the group convertible to an amino group, deprotecting the protected amino group or converting said convertible group to an amino group, and (3) is further required, converting the product to an acid addition salt.

The present study was designed to investigate the effects of the
 following factors on the perceived effort of the shoulder and
 elbow:

This invention relates to novel Amplifiable esters, processes for their production, and to an antibacterial agent comprising such as Amplifiable ester.

[illegible][illegible]

continued on p. 2.

BSPP:

It is clear that the Ampicillin esters of the invention have a low toxicity and longer duration of action, etc.

BSPP:

Experiments 1 to 3 are described below for demonstrating these advantages of the Ampicillin esters of the invention.

BSPP:

Each of the test compounds was orally administered to a 150 g. 10 day old, unstarved, Ampicillin free, 10 week, female rat, body weight of 200 g., 100 per cent body weight having been gained last overnight. The amount is equivalent to 100 mg. of a normal aqueous solution of Ampicillin. The blood was taken from the experimental animals periodically, and the concentration of Ampicillin in the serum was measured by a bioassay method. The blood Ampicillin level ratio was calculated from the following equation. **B/V**

BSPP:

The results given in Table 1 clearly show that the compounds of the invention show a high blood Ampicillin level over a longer period of time than the known penicillin ester 1.

BSPP:

Prodrugs such as Ampicillin bivaloxymethyl ester or Ampicillin phthaloyl ester have been known as orally administrable Ampicillin. The ester group of the Ampicillin ester of the invention (i.e., (2-oxo-1,3-dioxolan-4-yl)methyl group) is shown by a formula below in comparison with those of the known prodrugs.

BSPP:

It is clear therefore that the ester group of the Ampicillin ester of the invention quite differs from those of the known Ampicillin esters. It is surprising that the Ampicillin esters of the present invention have the aforesaid excellent properties as pharmaceuticals over these known Ampicillin esters.

BSPP:

According to one process of the invention, the Ampicillin ester or its acid addition salt of the invention can be produced by reacting a compound of the general formula **STR8** wherein A represents a protected amino group or a group convertible to an amino group, or its salt at the carboxyl group with a compound of the general formula **STR9** wherein R.sub.1 and R.sub.2 are as defined above, and X represents a halogen atom, and if required, when the resulting compound has the protected amino group or the group convertible to an amino group, eliminating the protecting group from the protected amino group or converting said convertible group to an amino group, and if further required, converting the product to its acid addition salt.

BSPP:

A compound corresponding to general formula II' in which A is a free amino group is a compound well known as penicillin and readily available commercially.

BSPP:

Analogously, the compound of general formula III can be produced by converting the free amino group of Ampicillin to the group A (in this case, the group A is desirably a protected amino group).

BSPP:

The compound of general formula II' can also be produced by reacting o-aminophenylsuccinic acid or its salt at the carboxyl group with a carboxylic acid of the general formula **STR10** wherein A is as defined hereinabove, or its acid addition salt at the methyl group. Such a process is the production of

wherein R₁ and R₂ are as defined, for example, in U.S. Pat. No. 3,711,714.

BBB:

Thus, the Ampicillin ester of general formula I, or its acid addition salt is formed. The acid addition salt is prepared by reacting the Ampicillin ester having a free amino group of general formula I with an acid, for example, an inorganic acid such as hydrochloric acid, hydrobromic acid, hydriodic acid and sulfuric acid, or an organic acid such as citric acid or tartaric acid.

BBB:

According to preferred embodiments of the process of the invention, there are provided a process for producing the ampicillin ester of general formula I or its acid addition salt, which comprises reacting a compound of general formula II, in which A is a substituent group or an amine group with the compound of general formula III, and converting the substituent group or the amine group of the product to an amino group, and thereafter if required, converting the product to its acid addition salt; and a process for producing a mineral acid salt (e.g., hydrochloride) of the ampicillin ester of general formula I which comprises reacting a compound of general formula (II) in which A is an amino group in the form of a mineral acid such as hydrochloride, with the compound of general formula (III).

BBB:

According to another process provided by the invention, the Ampicillin ester of general formula I, or its acid addition salt can be produced by reacting a compound of general formula ##STR13## wherein R.sub.1 and R.sub.2 are as defined above, or its acid addition salt, with a carboxylic acid of the general formula ##STR14## wherein A is as defined above, or its reactive derivative at the carboxyl group; thereafter, if required, when the resulting compound has the protected amino group or the group convertible to an amino group, eliminating the protective group from the protected amino group, or converting said convertible group to an amino group, and if further required, converting the product to an acid addition salt thereof.

BBB:

The compound of general formula (IV) can be produced by reacting D-aminopenicillanic acid or its salt at the carboxyl group with the compound of general formula (III); or by reacting D-protected aminopenicillanic acid or its salt at the carboxyl group with the compound of general formula (III) and then converting the protected amino group of the reaction product to an amino group.

BBB:

The former can be performed preferably by reacting D-aminopenicillanic acid or its salt at the carboxyl group with an equimolar amount, or a molar excess, of the compound of general formula III in an inert organic solvent such as tetrahydrofuran, dioxane or acetone in the optional presence of a base when D-aminopenicillanic acid is used, the presence of a base is preferred at a temperature of from about 0 degree C. to room temperature.

BBB:

The latter can be performed preferably by reacting D-protected aminopenicillanic acid such as D-aminopenicillanic acid having the amino group at the C-6 position protected with an acyl group or trityl group, or triaminopenicillanic acid having the amino group at the C-6 position protected as a Schiff base, or its salt at the carboxyl group, for example D-phenylacetylaminopenicillanic acid (benzylpenicillin), with the compound of general formula (III) under the same conditions as in the first-mentioned process, thereafter reacting the resulting D-protected aminopenicillanic acid ester with phosphorus pentachloride and a lower alcohol such as methanol at the temperature of dry ice-temperature in the presence of a basic compound such as triethylamine, pyridine and triethylamine, and thereafter removing water to give the resulting compound of general formula I.

EMER:

The reaction between the compound of general formula (I) or its acid addition salt and the compound of general formula (II) is a reversible reaction between the compound of general formula (II) and the compound of general formula (I) and a protecting group (R¹) is a group convertible to an amino group, the protecting group is removed from the protected amino group, or the convertible group is converted to an amino group and if desired, the product is converted to its acid addition salt. Thus, the Ampicillin ester of general formula (I) or its acid addition salt is formed.

EMER:

According to preferred embodiments of the above process, there are provided a process for producing the Ampicillin ester of general formula (I) or its acid addition salt which comprises reacting a compound of general formula (II) in which A is a Schiff base group or an amine group with the compound of general formula (I), thereafter converting the Schiff base group or the amine group (A) of the resulting compound to an amino group and if required, converting the product into its acid addition salt; and a process for producing an acid addition salt, such as a hydrochloride, of the Ampicillin ester of general formula (I) which comprises reacting a compound of general formula (II) in which A is in the form of an acid addition salt such as a hydrochloride with the compound of general formula (I).

EMER:

After the reaction, the Ampicillin of general formula (I) or its acid addition salt can be isolated and purified in a customary manner.

ESPER:

The Ampicillin ester of general formula (I) or its pharmaceutically acceptable acid addition salt is converted back to Ampicillin in vivo when administered to an animal. Accordingly, this invention also provides an antibacterial agent comprising the Ampicillin ester of general formula (I) or its pharmaceutically acceptable acid addition salt as an active ingredient.

EMER:

The antibacterial agent of this invention may consist only of the Ampicillin ester of general formula (I) or its pharmaceutically acceptable acid addition salt, or a mixture of it with a pharmaceutically acceptable carrier.

ESPER:

The pharmaceutically acceptable carrier may be those carriers which can be used in formulating Ampicillin. Examples are starch, lactose, hydroxypropyl cellulose, crystalline cellulose, magnesium stearate, and calcium stearate.

ESPER:

The antibacterial agent of this invention is administered to man and other animals in a dose of 1 to 20 mg/kg body weight (as calculated as the Ampicillin ester of general formula (I) or its pharmaceutically acceptable salt).

ESPER:

Ampicillin 2-methyl-2-oxo-1,3-dioxolen-4-yl methyl ester R.sub.1 = methyl, R.sub.2 = hydrogen,

EMER:

Ampicillin 2-oxo-1,3-dioxolen-4-yl methyl ester R.sub.1 and R.sub.2 = hydrogen,

ESPER:

Ampicillin 2-oxo-2-phenyl-1,3-dioxolen-4-yl methyl ester R.sub.1 = phenyl, R.sub.2 = hydrogen,

EMER:

Ampicillin 2,6-di-tert-butyl-xy-3-cyclohexen-1-yl ester R.sub.1 and R.sub.2 = tert-butyl or the group --R.sub.3--R.sub.3, and

BESTL:

1. Ampicillin 3-methyl-2-oxo-1,3-dioxolan-4-yl methyl ester hydrochloride compounds of the invention.

BESTL:

A. Ampicillin 3-methyl-2-oxo-1,3-dioxolan-4-yl methyl ester hydrochloride compounds of the invention.

BESTL:

B. Ampicillin 3-methyl-2-oxo-1,3-dioxolan-4-yl methyl ester hydrochloride compounds of the invention.

BESTL:

C. Ampicillin 3-methyl-2-oxo-1,3-dioxolan-4-yl methyl ester hydrochloride compound used as a control; see Section 1.1. B. 1, 2, 3, 4, 5.

BESTL:

D. Ampicillin trihydrate control.

BESTL:

TABLE 1
Last time of oral dosing: min. Item Ampicillin level ratio
Compounds A 2.8 2.9 2.1 1.8 1.8 1.3 of
the invention B 2.4 1.7 2.4 1.4 1.2 1. Known compound C 1.3 1.8 1.4 1.1 1.9
1.3 Control compound D 1.1 1.0 1.0 1.0 1.0 1.0

BESTL:

Ester group
Ampicillin pivaloxyloxy methyl ester
STR5 Ampicillin phthalidyl ester **STR6** Ampicillin ester of the
invention **STR7**

BESTL:

Ampicillin trihydrate 50 mg was dispersed in 6 ml of dimethyl formamide, and 100 mg of potassium hydrogen carbonate was added. The mixture was cooled to 0 degree. C. and stirred. Benzaldehyde 1.01 ml was added, and the mixture was stirred at 0 degree. C. for 1.8 hours. Then, 125 mg of potassium hydrogen carbonate and 320 mg of 4-bromomethyl-3-phenyl-1,3-dioxolan-2-one were added, and the mixture was further stirred at 0 degree. C. for 3 hours.

BESTL:

Water 10 ml was added, and the mixture was concentrated under reduced pressure to 10 ml or less. The aqueous layer was repeatedly washed with ethyl acetate, and saturated with sodium chloride. The separated oily substance was extracted with 50 ml of methylene chloride, washed with a saturated aqueous solution of sodium chloride and dried over anhydrous sodium sulfate. The dried organic layer was concentrated until the amount of methylene chloride decreased to one half. Isopropyl alcohol (30 ml) was added, and the mixture was again concentrated under reduced pressure to give a colorless solid. The solid was collected by filtration, and washed successively with isopropyl alcohol and ether to give 320 mg (yield 46.4%) of Ampicillin 3-methyl-2-oxo-1,3-dioxolan-4-yl methyl ester hydrochloride as a colorless solid.

BESTL:

The resulting Ampicillin ester hydrochloride was incubated in 4% mouse blood in pH 7.4 phosphate buffer at 37 degree. C. for 15 minutes, and then subjected to bioautography. It was found to be completely converted to Ampicillin.

BESTL:

Ampicillin trihydrate 50 mg was dispersed in 6 ml of dimethyl formamide, and 100 mg of potassium hydrogen carbonate was added. The mixture was cooled to 0 degree. C. and stirred. Benzaldehyde 1.01 ml was added, and the mixture was stirred at 0 degree. C. for 1.8 hours. Then, 125 mg of potassium hydrogen carbonate and

EXPER:

10-grams of 2-oxo-1,3-dioxolen-4-ylmethyl d-aminopenicillanate hydrochloride was suspended in 100 ml of methylene chloride. The mixture was stirred at 0.degree. C., and by passing nitrogen gas, the sym. chloride of the penicillanate was formed. Ethylmagnesium bromide (20 g) was added, and the mixture was stirred at 0.degree. C. for 4 hours. The solid precipitated was collected by filtration, and repeatedly washed with methylene chloride to give 13.1 g (yield 90%) of 2-oxo-1,3-dioxolen-4-ylmethyl ester hydrochloride as a colorless amorphous solid.

EXPER:

10-grams of 2-oxo-1,3-dioxolen-4-ylmethyl d-aminopenicillanate hydrochloride was suspended in 100 ml of methylene chloride, and by passing nitrogen gas, the sym. chloride was formed. The mixture was stirred at 0.degree. C. for 4 hours. Then, 10 ml of 10% aqueous sodium hydroxide solution was added, and the mixture was stirred for 2 hours and then for another 2 hours at room temperature.

EXPER:

After the reaction, the solid was separated by filtration, and the filtrate was concentrated under reduced pressure. The resulting syrup was dissolved in water, and washed with ethyl acetate. The aqueous layer was saturated with sodium chloride, and the separated oily substance was extracted with methylene chloride. The extract was washed with a saturated aqueous solution of sodium chloride and concentrated until the amount of methylene chloride decreased to half. Upon addition of isopropyl alcohol, a colorless solid was precipitated. The solid was collected by filtration and washed with isopropyl alcohol and ether to give 132 mg (yield 54%) of Ampicillin (5-methyl-2-oxo-1,3-dioxolen-4-yl)methyl ester hydrochloride as an amorphous solid.

EXPER:

From 1.0 g of the resulting ester hydrochloride and 95 mg of 2-oxo-1,3-dioxolen-4-ylmethyl d-aminopenicillanate hydrochloride, 145 mg (yield 56%) of Ampicillin (5-methyl-2-oxo-1,3-dioxolen-4-yl)methyl ester hydrochloride was obtained as a colorless amorphous solid.

EXPER:

Five grams of 5-methyl-2-oxo-1,3-dioxolen-4-ylmethyl d-aminopenicillanate p-toluenesulfonate was suspended in 300 ml of ethyl acetate. To the suspension was added at 0.degree. C. 100 ml of a 10% aqueous solution of sodium hydrogen carbonate cooled with ice. The mixture was vigorously stirred. The ethyl acetate layer was separated, washed with ice water, dried at 0.degree. C. over anhydrous magnesium sulfate, and concentrated under reduced pressure to give a pale yellow syrup. The syrup was dissolved in 50 ml of methylene chloride. The solution was cooled to 0.degree. C., and 1 g of potassium hydrogen carbonate and 2.1 g of 2-oxo-1,3-dioxolen-4-ylmethyl d-aminopenicillanate hydrochloride were added, and the mixture was stirred at 0.degree. C. for 4 hours. After the reaction, the insoluble material was separated by filtration, and the filtrate was concentrated under reduced pressure. The resulting syrup was dissolved in water and washed with ethyl acetate. The aqueous layer was saturated with sodium chloride. The separated oily substance was extracted with methylene chloride, washed with a saturated aqueous solution of sodium chloride and dried over anhydrous magnesium sulfate. The dried solution was concentrated under reduced pressure until the amount of methylene chloride decreased to one half. Isopropyl alcohol was added, and the mixture was again concentrated under reduced pressure to give a colorless solid. The solid was collected by filtration, and washed with ether to give 1.1 g (yield 61%) of Ampicillin (5-methyl-2-oxo-1,3-dioxolen-4-yl)methyl ester hydrochloride as a colorless amorphous solid.

EXPER:

An ethanol solution of the hydroxypropyl cellulose was prepared and added to the Ampicillin ester hydrochloride and lactose. They were kneaded, extruded through a screen, and dried in a vacuum oven.

THER:

1. An α -amylase ester of the formula $\text{H}_2\text{N}(\text{CH}_2)_n\text{COO}(\text{R})$ wherein R represents a hydrocarbon, a methyl group or an aryl group, and n represents a hydrocarbon chain may be taken together with $\text{H}_2\text{N}(\text{CH}_2)_m\text{COO}(\text{R})$ to form a divalent chain residue, or a pharmaceutically acceptable acid addition salt thereof.

THER:

2. Production of Ampicillin (2-oxo-5-phenyl-1,3-dioxolen-4-yl)methyl ester hydrochloride

THER:

3. Production of Ampicillin (3-methyl-2-oxo-1,3-dioxolen-4-yl)methyl ester hydrochloride

THER:

4. Production of Ampicillin (2,6-dihydroxydioxo-4-phenylhexenyl) ester hydrochloride

THER:

5. Production of Ampicillin (2-oxo-1,3-dioxolen-4-yl)methyl ester hydrochloride

THER:

6. Production of Ampicillin (5-methyl-2-oxo-1,3-dioxolen-4-yl)methyl ester hydrochloride

THER:

7. Production of Ampicillin (2-oxo-5-phenyl-1,3-dioxolen-4-yl)methyl ester hydrochloride

THER:

8. Production of Ampicillin (5-methyl-2-oxo-1,3-dioxolen-4-yl)methyl ester hydrochloride

THER:

9. Production of Ampicillin (5-methyl-2-oxo-1,3-dioxolen-4-yl)methyl ester hydrochloride 356.7 mg Lactose 36.3 mg Magnesium stearate 5.0 mg 400 mg in total

THER:

10. Production of Ampicillin (5-methyl-2-oxo-1,3-dioxolen-4-yl)methyl ester hydrochloride 356.7 mg Lactose 36.3 mg Magnesium stearate 5.0 mg 400 mg in total

THER:

11. Production of Ampicillin (5-methyl-2-oxo-1,3-dioxolen-4-yl)methyl ester hydrochloride 356.7 mg Lactose 36.3 mg Magnesium stearate 5.0 mg 400 mg in total

THER:

12. Production of Ampicillin (5-methyl-2-oxo-1,3-dioxolen-4-yl)methyl ester hydrochloride 356.7 mg Lactose 36.3 mg Magnesium stearate 5.0 mg 400 mg in total

THER:

13. An α -amylase ester of the formula $\text{H}_2\text{N}(\text{CH}_2)_n\text{COO}(\text{R})$ wherein R represents a hydrocarbon, a methyl group or an aryl group, and n represents a hydrocarbon chain may be taken together with $\text{H}_2\text{N}(\text{CH}_2)_m\text{COO}(\text{R})$ to form a divalent chain residue, or a pharmaceutically acceptable acid addition salt thereof.

THER:

14. An antibacterial agent comprising an antibacterially effective amount of an Ampicillin ester of the formula $\text{H}_2\text{N}(\text{CH}_2)_n\text{COO}(\text{R})$ wherein R represents a hydrocarbon, a methyl group or an aryl group, and n represents a hydrocarbon chain may be taken together with $\text{H}_2\text{N}(\text{CH}_2)_m\text{COO}(\text{R})$ to form a divalent chain residue, or a pharmaceutically acceptable acid addition salt thereof.

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1. $\frac{1}{2} \times \frac{1}{2} = \frac{1}{4}$ (Probability of getting two heads)

Many, semisynthetic beta-lactam derivatives such as ampicillin, amoxycillin, cefaclor, cephalexin, cephadroxil and cephalexoglycin are, on an industrial scale, prepared by chemical methods, for example by reacting an amine beta-lactam such as D-phenylpenicillanic acid, usually having its carboxyl group protected, with an activated side chain derivative, followed by the removal of the protecting group by hydrolysis. For example, ampicillin (2-(S)-alpha-aminophenylacetamido-penicillanic acid) can be prepared by reacting D-phenylpenicillin, having a suitably protected carboxyl group, with phenylglycylglycine or D-phenylglycine, followed by removal of the protecting group by hydrolysis. These reactions typically involve reacting with an activated carboxylic anhydride and organic solvents like acetonitrile and dimethyl sulfoxide.

Within the last years, there has been an increasing amount of publications concerning the possibility of using the preparation of penicillins and cephalosporins by a method of acylation, i.e. by the reaction of an acyl chloride with a penicillin, which is well known in the prior art. For example, the reaction of penicillin G with 2-aminocaprylic acid is known from West German patent application having publication No. 1,168,190, Austrian Patent No. 248,446, Dutch patent application No. 72-19138, West German patent application having publication No. 2,441,616, European patent application having publication No. 338,781 and British application having publication No. 1,311,311.

Independently, it has now been found that improved process conditions are obtained if the enzymatic acylation of the amino (beta)-lactam is performed at substantially equal concentrations of both the parent amino (beta)-lactam and the corresponding acylating agent. Preferably the concentrations of the parent amino (beta)-lactam and of the corresponding acylating agent are chosen such that the parent amino (beta)-lactam is at the lowest value of 1 in the estimated molar ratio of the parent amino (beta)-lactam and of the estimated concentration of the corresponding acylating agent.

DEPR:

It has now, surprisingly, been found that improved process conditions are obtained if the enzymatic acylation of the amino (beta)-lactam is performed at substantially equal concentrations of both the parent amino (beta)-lactam and the corresponding acylating agent. Preferably the concentrations of the parent amino (beta)-lactam and of the corresponding acylating agent are chosen such that the parent amino (beta)-lactam is at the lowest value of 1 in the estimated molar ratio of the parent amino (beta)-lactam and of the estimated concentration of the corresponding acylating agent.

DEPR:

The (beta)-lactam derivative formed may precipitate during the reaction and, also, the acid form of the acylating agent such as L-phenylglycine and L-p-hydroxyphenylglycine may precipitate. Hence, in some cases the reaction mixture will be a slurry throughout the reaction.

DEPR:

The parent amino (beta)-lactam has a free amino group which is acylated by the reaction according to this invention. The amino (beta)-lactam may conveniently be 6-APA, 7-ADCA, 7-ACA or 7-ACCC.

DEPR:

The amino (beta)-lactam, for example 6-APA or 7-ADCA, used in the process of this invention may be obtained by enzymatic hydrolysis of the fermented penicillins or cephalosporins, for example penicillin V, penicillin G or cephalosporin C or their ring enlarged analogues (for example V-ACA and G-ACA) or derivatives thereof followed by removal of the hydrolysis by-product, if desired (phenoxycetic acid etc.). Advantageously, the crude solution can be used directly without further purification or dilution.

DEPR:

The acylating agent may be in an activated form. Preferably, the acylating agent is an amide or an ester. The acylating agent may be a derivative of L-phenylglycine, D-p-hydroxyphenylglycine, D-2,5-dihydroxyphenylglycine or mandelic acid, such as a lower alkyl ester (methyl, ethyl, n-propyl or isopropyl ester) or an amide which is unsubstituted or substituted in the α -position, e.g. the acylating agent may be used in the form of a salt, for example, the sulphate salt or the H-sulphate salt. The acylating agent may be added in an active form or the active form may be formed in situ.

DEPR:

The solubility of the acylating agent such as the L-phenylglycine or L-p-hydroxyphenylglycine derivative will vary with the identity of the derivative and with the composition of the reaction medium. In an aqueous system as used in the examples, the solubility of the hydrochloride salt of L-phenylglycine amide is typically approximately 40 mM. However, the solubility is not dependent on the salt components in the solution, as well as on the pH value and the temperature of the solution. As a further example, the solubility of the sulphate form of D-phenylglycine amide is about 3.3 M within a pH range from 2.5 to 6.5.

DEPR:

Examples of (beta)-lactam derivatives that may be prepared by the process of this invention are amoxicillin, cefadixor, cephalexin, cephadraxil, cephadrone, cephadrin and cefmandolil.

DEPR:

The enzyme to be used in the process of this invention may be any enzyme catalysing the reaction in question. Such enzymes have been known since at least 1961. Enzymes to be used are, for example, termed penicillin aminase or penicillin acylase and classified as E.C. 3.5.1.11. A number of microbial

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361	362	363	364	365	366
367	368	369	370	371	372
373	374	375	376	377	378
379	380	381	382	383	384
385	386	387	388	389	390
391	392	393	394	395	396
397	398	399	400	401	402
403	404	405	406	407	408
409	410	411	412	413	

The suitable pH value depends, inter alia, on the type of the enzyme used. For Proteinase with enzyme, the pH value is typically in the range from 7 to 9, preferably in the range from 8.1 to 9. For the preparation of an emulsion, a pH value in the range from 7.8 to 8.4 is preferred. Control of the pH value may be used.

2000	2001	2002	2003	2004	2005
1.0	1.0	1.0	1.0	1.0	1.0
1.0	1.0	1.0	1.0	1.0	1.0
1.0	1.0	1.0	1.0	1.0	1.0

$$Z_1 = \frac{1}{\sqrt{2}} \begin{pmatrix} 1 & 1 \\ 1 & -1 \end{pmatrix} \Rightarrow Z_1^{-1} = \frac{1}{2} \begin{pmatrix} 1 & 1 \\ 1 & -1 \end{pmatrix} \Rightarrow Z_1^{-1} \begin{pmatrix} 1 & 0 \\ 0 & 1 \end{pmatrix} Z_1 = \frac{1}{2} \begin{pmatrix} 1 & 1 \\ 1 & -1 \end{pmatrix} \begin{pmatrix} 1 & 0 \\ 0 & 1 \end{pmatrix} \begin{pmatrix} 1 & 1 \\ 1 & -1 \end{pmatrix} = \frac{1}{2} \begin{pmatrix} 1 & 1 \\ 1 & -1 \end{pmatrix} \begin{pmatrix} 1 & 1 \\ 1 & -1 \end{pmatrix} = \frac{1}{2} \begin{pmatrix} 2 & 0 \\ 0 & 2 \end{pmatrix} = \begin{pmatrix} 1 & 0 \\ 0 & 1 \end{pmatrix}$$
[illegible]

Using the process according to this invention, an extraordinary high molar ratio between the amount of olefin-ligand derivative which can be recovered and the total amount of olefin and the resulting agent can be obtained. These high ratios are obtained using the technique of this invention and properly controlling the concentration of the resulting agent, the ratio between the concentration of resulting agent and the starting olefin-ligand, the pH value and the ligand. Thus, a ratio of 1.4 was obtained in Example 1 below using the process according to the present invention. In a comparative batch process, vide Example 1 below, a molar ratio of only 1.4 was obtained. In addition, the yields of isolated product obtained in this example were 40 and 80, respectively.

1. 2. 3. 4. 5. 6. 7. 8. 9. 10. 11. 12. 13. 14. 15. 16. 17. 18. 19. 20. 21. 22. 23. 24. 25. 26. 27. 28. 29. 30. 31. 32. 33. 34. 35. 36. 37. 38. 39. 40. 41. 42. 43. 44. 45. 46. 47. 48. 49. 50. 51. 52. 53. 54. 55. 56. 57. 58. 59. 60. 61. 62. 63. 64. 65. 66. 67. 68. 69. 70. 71. 72. 73. 74. 75. 76. 77. 78. 79. 80. 81. 82. 83. 84. 85. 86. 87. 88. 89. 90. 91. 92. 93. 94. 95. 96. 97. 98. 99. 100. 101. 102. 103. 104. 105. 106. 107. 108. 109. 110. 111. 112. 113. 114. 115. 116. 117. 118. 119. 120. 121. 122. 123. 124. 125. 126. 127. 128. 129. 130. 131. 132. 133. 134. 135. 136. 137. 138. 139. 140. 141. 142. 143. 144. 145. 146. 147. 148. 149. 150. 151. 152. 153. 154. 155. 156. 157. 158. 159. 160. 161. 162. 163. 164. 165. 166. 167. 168. 169. 170. 171. 172. 173. 174. 175. 176. 177. 178. 179. 180. 181. 182. 183. 184. 185. 186. 187. 188. 189. 190. 191. 192. 193. 194. 195. 196. 197. 198. 199. 200. 201. 202. 203. 204. 205. 206. 207. 208. 209. 210. 211. 212. 213. 214. 215. 216. 217. 218. 219. 220. 221. 222. 223. 224. 225. 226. 227. 228. 229. 230. 231. 232. 233. 234. 235. 236. 237. 238. 239. 240. 241. 242. 243. 244. 245. 246. 247. 248. 249. 250. 251. 252. 253. 254. 255. 256. 257. 258. 259. 260. 261. 262. 263. 264. 265. 266. 267. 268. 269. 270. 271. 272. 273. 274. 275. 276. 277. 278. 279. 280. 281. 282. 283. 284. 285. 286. 287. 288. 289. 290. 291. 292. 293. 294. 295. 296. 297. 298. 299. 300. 301. 302. 303. 304. 305. 306. 307. 308. 309. 310. 311. 312. 313. 314. 315. 316. 317. 318. 319. 320. 321. 322. 323. 324. 325. 326. 327. 328. 329. 330. 331. 332. 333. 334. 335. 336. 337. 338. 339. 340. 341. 342. 343. 344. 345. 346. 347. 348. 349. 350. 351. 352. 353. 354. 355. 356. 357. 358. 359. 360. 361. 362. 363. 364. 365. 366. 367. 368. 369. 370. 371. 372. 373. 374. 375. 376. 377. 378. 379. 380. 381. 382. 383. 384. 385. 386. 387. 388. 389. 390. 391. 392. 393. 394. 395. 396. 397. 398. 399. 400. 401. 402. 403. 404. 405. 406. 407. 408. 409. 410. 411. 412. 413. 414. 415. 416. 417. 418. 419. 420. 421. 422. 423. 424. 425. 426. 427. 428. 429. 430. 431. 432. 433. 434. 435. 436. 437. 438. 439. 440. 441. 442. 443. 444. 445. 446. 447. 448. 449. 450. 451. 452. 453. 454. 455. 456. 457. 458. 459. 460. 461. 462. 463. 464. 465. 466. 467. 468. 469. 470. 471. 472. 473. 474. 475. 476. 477. 478. 479. 480. 481. 482. 483. 484. 485. 486. 487. 488. 489. 490. 491. 492. 493. 494. 495. 496. 497. 498. 499. 500. 501. 502. 503. 504. 505. 506. 507. 508. 509. 510. 511. 512. 513. 514. 515. 516. 517. 518. 519. 520. 521. 522. 523. 524. 525. 526. 527. 528. 529. 530. 531. 532. 533. 534. 535. 536. 537. 538. 539. 540. 541. 542. 543. 544. 545. 546. 547. 548. 549. 550. 551. 552. 553. 554. 555. 556. 557. 558. 559. 560. 561. 562. 563. 564. 565. 566. 567. 568. 569. 570. 571. 572. 573. 574. 575. 576. 577. 578. 579. 580. 581. 582. 583. 584. 585. 586. 587. 588. 589. 590. 591. 592. 593. 594. 595. 596. 597. 598. 599. 600. 601. 602. 603. 604. 605. 606. 607. 608. 609. 610. 611. 612. 613. 614. 615. 616. 617. 618. 619. 620. 621. 622. 623. 624. 625. 626. 627. 628. 629. 630. 631. 632. 633. 634. 635. 636. 637. 638. 639. 640. 641. 642. 643. 644. 645. 646. 647. 648. 649. 650. 651. 652. 653. 654. 655. 656. 657. 658. 659. 660. 661. 662. 663. 664. 665. 666. 667. 668. 669. 670. 671. 672. 673. 674. 675. 676. 677. 678. 679. 680. 681. 682. 683. 684. 685. 686. 687. 688. 689. 690. 691. 692. 693. 694. 695. 696. 697. 698. 699. 700. 701. 702. 703. 704. 705. 706. 707. 708. 709. 710. 711. 712. 713. 714. 715. 716. 717. 718. 719. 720. 721. 722. 723. 724. 725. 726. 727. 728. 729. 730. 731. 732. 733. 734. 735. 736. 737. 738. 739. 740. 741. 742. 743. 744. 745. 746. 747. 748. 749. 750. 751. 752. 753. 754. 755. 756. 757. 758. 759. 760. 761. 762. 763. 764. 765. 766. 767. 768. 769. 770. 771. 772. 773. 774. 775. 776. 777. 778. 779. 780. 781. 782. 783. 784. 785. 786. 787. 788. 789. 790. 791. 792. 793. 794. 795. 796. 797. 798. 799. 800. 801. 802. 803. 804. 805. 806. 807. 808. 809. 810. 811. 812. 813. 814. 815. 816. 817. 818. 819. 820. 821. 822. 823. 824. 825. 826. 827. 828. 829. 830. 831. 832. 833. 834. 835. 836. 837. 838. 839. 840. 84

1000

1818:

An estimation of penicillin G acylase activity the following is used: the unit corresponds to the amount of amoxicillin that hydrolyzes per minute in 1 mmole penicillin G under standard conditions: 1.0 g/l, eq. 1 Penicillin G potassium salt, 0.1 M potassium phosphate buffer, pH value 6.1, 20.degree. C.

1819:

The equipment for this experiment consisted of (see FIG. 1) a thermostated reactor having a volume of 1.5 liter, equipped with a three-bladed impeller and a cover with 40 mm in diameter open area at the top. The reactor was connected to an agitator system using 4 M sodium hydroxide as titrant. A valve was positioned at the outlet of the reactor. The outlet of the valve was connected via a pump to a basket centrifuge equipped with a polypropylene bag having a capacity of 1-6 l/min. The outlet from the centrifuge was connected via a pump to a feed tank equipped with a stirrer and a glass sinter bottom. The outlet from the feed tank was connected via a pump to the reactor.

1820:

A mixture consisting of D-HPGM (21.7 g, 120 mmol) and L-ASA (43.3 g, 120 mmol) in 75 ml water was added to the reactor with the bottom valve closed. The stirring was started. Immobilized Penicillin G acylase (200 g/l, size 40-60 µm) made up to 200 ml was added to the reactor. The pH value was maintained at 6.1. The reaction temperature was about 20.degree. C. Under those conditions, the reaction mixture was almost saturated with D-HPGM and L-ASA. Then, the bottom valve was opened allowing the reaction mixture from the reactor to enter the centrifuge. Thereafter, the mother liquor from the centrifuge was pumped into the feed tank wherein D-HPGM (21.7 g, 120 mmol) and L-ASA (43.3 g, 120 mmol) were loaded. The total volume of the suspension in the feed tank was kept at about 75 ml. A flow of about 100 ml/min was maintained in the system. The concentration of reaction components in the reactor, in the reactor outlet, in the centrifuge outlet, in the feed tank and in the feed tank outlet were monitored by analytical HPLC.

1821:

As the reaction proceeded, the amoxicillin and D-HPG formed started to precipitate out of solution. The crystals were separated from the immobilized enzyme particles by the bottom sieve in the reactor and the crystal suspension was led to the centrifuge where the crystals were separated from the mother liquor. When the mother liquor, which was now almost saturated with respect to D-HPGM and L-ASA, passed through the feed tank, some of the solid D-HPGM and L-ASA present dissolved such that the outlet from the feed tank contained saturated D-HPGM and L-ASA. When the total concentrations of D-HPGM and L-ASA came down to about 225 mM HPGM and 225 mM L-ASA, more solid substrate was added to the feed tank. At intervals the crystals in the centrifuge were washed with water, the washing liquid was added to the reactor. The amount of water used was sufficient to keep the volume in the reactor at its starting level. When washed crystals were separated in the centrifuge, the centrifuge was emptied.

1822:

After about 12 hours, the dosing of D-HPGM and L-ASA was stopped. After 14 hours, the L-ASA concentration reached 1. mM and the reaction was stopped. The amounts of reaction components are given in Table 1, below.

1823:

A comparative experiment was performed at batchwise conditions and the reaction was stopped at the moment where the optimum yield of amoxicillin was obtained (1.4 mmol). The reaction temperature was about 20.degree. C., the pH value was about 6.1 and the amount of the enzyme, mentioned in this example above, was used. The total volume of the reaction mixture was 1 liter. The reaction was performed in the reactor mentioned above. After 7 hours the mother liquor was poured, and the crystals were separated from the immobilized enzyme particles by the bottom sieve in the reactor. The crystal suspension was filtered. The amounts of reaction components are given in Table 1, below.

[illegible]

A mixture consisting of 1-HBA (2.0×10^{-3} mol) and 1-AA (1.0×10^{-3} mol) in 10 ml water, which was adjusted at a pH value of 4.1 by adding 4 M ammonium hydroxide, was placed in the reactor with the bottom valve closed. The stirring was started. Immobilized Penicillin G-lysozyme (2000 U, type 10-11, 100 mg) made by 10 ml of 10% was added to the reactor. The pH value was maintained at 4.1. The reaction temperature was at $37 \pm 0.1^\circ\text{C}$. After these conditions the reaction mixture was stirred at constant rate with 1-HBA and 1-AA. Then the bottom valve was opened and the reaction mixture in the feed tank after the immobilized lysozyme, the buffer liquid in the centrifuge was separated. The feed tank where-in 1-HBA (2.0×10^{-3} mol) and 1-AA (1.0×10^{-3} mol) were added. The total volume of the suspension in the feed tank was kept at about 10 ml. A flow of about 10 ml/min was maintained in the system. The concentration of reaction components in the reactor, in the reactor outlet, in the centrifuge outlet, in the feed tank, and in the feed tank outlet were maintained analytical HPLC.

[illegible]

The crystals formed in the reactor were separated from the immobilized enzyme solution by the mother liquor which was in the reactor. The crystal suspension was fed into the centrifuge where the crystals were separated from the mother liquor. The mother liquor was passed through the feed tank. When the total concentration in the feed tank came down to about 180 mM HPGA and 225 mM GA3, more solid was added to the feed tank. At intervals the crystals in the centrifuge were washed with water, the washing liquid was added to the reactor. The amount of water used was sufficient to keep the volume in the reactor at its starting level. After washing the crystals in the centrifuge, the centrifuge was turned.

[illegible]

After about 14 hours, the dosing of D-HPGA to the feed tank was stopped while the D-HPGA loading was maintained in order to keep D-HPGA at saturation. After 14 hours, the 6-AGA concentration reached 20 mM and the reaction was stopped. The amounts of reaction components are given in Table 3, below.

[illegible]

^a Values are means ± SD.

1. ☐ 2. ☐ 3. ☐ 4. ☐ 5. ☐ 6. ☐ 7. ☐ 8. ☐ 9. ☐ 10. ☐ 11. ☐ 12. ☐ 13. ☐ 14. ☐ 15. ☐ 16. ☐ 17. ☐ 18. ☐ 19. ☐ 20. ☐ 21. ☐ 22. ☐ 23. ☐ 24. ☐ 25. ☐ 26. ☐ 27. ☐ 28. ☐ 29. ☐ 30. ☐ 31. ☐ 32. ☐ 33. ☐ 34. ☐ 35. ☐ 36. ☐ 37. ☐ 38. ☐ 39. ☐ 40. ☐ 41. ☐ 42. ☐ 43. ☐ 44. ☐ 45. ☐ 46. ☐ 47. ☐ 48. ☐ 49. ☐ 50. ☐ 51. ☐ 52. ☐ 53. ☐ 54. ☐ 55. ☐ 56. ☐ 57. ☐ 58. ☐ 59. ☐ 60. ☐ 61. ☐ 62. ☐ 63. ☐ 64. ☐ 65. ☐ 66. ☐ 67. ☐ 68. ☐ 69. ☐ 70. ☐ 71. ☐ 72. ☐ 73. ☐ 74. ☐ 75. ☐ 76. ☐ 77. ☐ 78. ☐ 79. ☐ 80. ☐ 81. ☐ 82. ☐ 83. ☐ 84. ☐ 85. ☐ 86. ☐ 87. ☐ 88. ☐ 89. ☐ 90. ☐ 91. ☐ 92. ☐ 93. ☐ 94. ☐ 95. ☐ 96. ☐ 97. ☐ 98. ☐ 99. ☐ 100. ☐

1997, 1998, 1999, 2000, 2001, 2002, 2003, 2004, 2005, 2006, 2007, 2008, 2009, 2010, 2011, 2012, 2013, 2014, 2015, 2016, 2017, 2018, 2019, 2020, 2021, 2022, 2023, 2024, 2025, 2026, 2027, 2028, 2029, 2030, 2031, 2032, 2033, 2034, 2035, 2036, 2037, 2038, 2039, 2040, 2041, 2042, 2043, 2044, 2045, 2046, 2047, 2048, 2049, 2050, 2051, 2052, 2053, 2054, 2055, 2056, 2057, 2058, 2059, 2060, 2061, 2062, 2063, 2064, 2065, 2066, 2067, 2068, 2069, 2070, 2071, 2072, 2073, 2074, 2075, 2076, 2077, 2078, 2079, 2080, 2081, 2082, 2083, 2084, 2085, 2086, 2087, 2088, 2089, 2090, 2091, 2092, 2093, 2094, 2095, 2096, 2097, 2098, 2099, 2100, 2101, 2102, 2103, 2104, 2105, 2106, 2107, 2108, 2109, 2110, 2111, 2112, 2113, 2114, 2115, 2116, 2117, 2118, 2119, 2120, 2121, 2122, 2123, 2124, 2125, 2126, 2127, 2128, 2129, 2130, 2131, 2132, 2133, 2134, 2135, 2136, 2137, 2138, 2139, 2140, 2141, 2142, 2143, 2144, 2145, 2146, 2147, 2148, 2149, 2150, 2151, 2152, 2153, 2154, 2155, 2156, 2157, 2158, 2159, 2160, 2161, 2162, 2163, 2164, 2165, 2166, 2167, 2168, 2169, 2170, 2171, 2172, 2173, 2174, 2175, 2176, 2177, 2178, 2179, 2180, 2181, 2182, 2183, 2184, 2185, 2186, 2187, 2188, 2189, 2190, 2191, 2192, 2193, 2194, 2195, 2196, 2197, 2198, 2199, 2200, 2201, 2202, 2203, 2204, 2205, 2206, 2207, 2208, 2209, 2210, 2211, 2212, 2213, 2214, 2215, 2216, 2217, 2218, 2219, 2220, 2221, 2222, 2223, 2224, 2225, 2226, 2227, 2228, 2229, 2230, 2231, 2232, 2233, 2234, 2235, 2236, 2237, 2238, 2239, 2240, 2241, 2242, 2243, 2244, 2245, 2246, 2247, 2248, 2249, 2250, 2251, 2252, 2253, 2254, 2255, 2256, 2257, 2258, 2259, 2260, 2261, 2262, 2263, 2264, 2265, 2266, 2267, 2268, 2269, 2270, 2271, 2272, 2273, 2274, 2275, 2276, 2277, 2278, 2279, 2280, 2281, 2282, 2283, 2284, 2285, 2286, 2287, 2288, 2289, 2290, 2291, 2292, 2293, 2294, 2295, 2296, 2297, 2298, 2299, 2300, 2301, 2302, 2303, 2304, 2305, 2306, 2307, 2308, 2309, 2310, 2311, 2312, 2313, 2314, 2315, 2316, 2317, 2318, 2319, 2320, 2321, 2322, 2323, 2324, 2325, 2326, 2327, 2328, 2329, 2330, 2331, 2332, 2333, 2334, 2335, 2336, 2337, 2338, 2339, 2340, 2341, 2342, 2343, 2344, 2345, 2346, 2347, 2348, 2349, 2350, 2351, 2352, 2353, 2354, 2355, 2356, 2357, 2358, 2359, 2360, 2361, 2362, 2363, 2364, 2365, 2366, 2367, 2368, 2369, 2370, 2371, 2372, 2373, 2374, 2375, 2376, 2377, 2378, 2379, 2380, 2381, 2382, 2383, 2384, 2385, 2386, 2387, 2388, 2389, 2390, 2391, 2392, 2393, 2394, 2395, 2396, 2397, 2398, 2399, 2400, 2401, 2402, 2403, 2404, 2405, 2406, 2407, 2408, 2409, 2410, 2411, 2412, 2413, 2414, 2415, 2416, 2417, 2418, 2419, 2420, 2421, 2422, 2423, 2424, 2425, 2426, 2427, 2428, 2429, 2430, 2431, 2432, 2433, 2434, 2435, 2436, 2437, 2438, 2439, 2440, 2441, 2442, 2443, 2444, 2445, 2446, 2447, 2448, 2449, 2450, 2451, 2452, 2453, 2454, 2455, 2456, 2457, 2458, 2459, 2460, 2461, 2462, 2463, 2464, 2465, 2466, 2467, 2468, 2469, 2470, 2471, 2472, 2473, 2474, 2475, 2476, 2477, 2478, 2479, 2480, 2481, 2482, 2483, 2484, 2485, 2486, 2487, 2488, 2489, 2490, 2491, 2492, 2493, 2494, 2495, 2496, 2497, 2498, 2499, 2500, 2501, 2502, 2503, 2504, 2505, 2506, 2507, 2508, 2509, 2510, 2511, 2512, 2513, 2514, 2515, 2516, 2517, 2518, 2519, 2520, 2521, 2522, 2523, 2524, 2525, 2526, 2527, 2528, 2529, 2530, 2531, 2532, 2533, 2534, 2535, 2536, 2537, 2538, 2539, 2540, 2541, 2542, 2543, 2544, 2545, 2546, 2547, 2548, 2549, 2550, 2551, 2552, 2553, 2554, 2555, 2556, 2557, 2558, 2559, 2560, 2561, 2562, 2563, 2564, 2565, 2566, 2567, 2568, 2569, 2570, 2571, 2572, 2573, 2574, 2575, 2576, 2577, 2578, 2579, 2580, 2581, 2582, 2583, 2584, 2585, 2586, 2587, 2588, 2589, 2590, 2591, 2592, 2593, 2594, 2595, 2596, 2597, 2598, 2599, 2600, 2601, 2602, 2603, 2604, 2605, 2606, 2607, 2608, 2609, 2610, 2611, 2612, 2613, 2614, 2615, 2616, 2617, 2618, 2619, 2620, 2621, 2622, 2623, 2624, 2625, 2626, 2627, 2628, 2629, 2630, 2631, 2632, 2633, 2634, 2635, 2636, 2637, 2638, 2639, 2640, 2641, 2642, 2643, 2644, 2645, 2646, 2647, 2648, 2649, 2650, 2651, 2652, 2653, 2654, 2655, 2656, 2657, 2658, 2659, 2660, 2661, 2662, 2663, 2664, 2665, 2666, 2667, 2668, 2669, 2670, 2671, 2672, 2673, 2674, 2675, 2676, 2677, 2678, 26

1994, 1995, 1996, 1997, 1998, 1999, 2000, 2001, 2002, 2003, 2004, 2005, 2006, 2007, 2008, 2009, 2010, 2011, 2012, 2013, 2014, 2015, 2016, 2017, 2018, 2019, 2020, 2021, 2022, 2023, 2024, 2025, 2026, 2027, 2028, 2029, 2030, 2031, 2032, 2033, 2034, 2035, 2036, 2037, 2038, 2039, 2040, 2041, 2042, 2043, 2044, 2045, 2046, 2047, 2048, 2049, 2050, 2051, 2052, 2053, 2054, 2055, 2056, 2057, 2058, 2059, 2060, 2061, 2062, 2063, 2064, 2065, 2066, 2067, 2068, 2069, 2070, 2071, 2072, 2073, 2074, 2075, 2076, 2077, 2078, 2079, 2080, 2081, 2082, 2083, 2084, 2085, 2086, 2087, 2088, 2089, 2090, 2091, 2092, 2093, 2094, 2095, 2096, 2097, 2098, 2099, 2100, 2101, 2102, 2103, 2104, 2105, 2106, 2107, 2108, 2109, 2110, 2111, 2112, 2113, 2114, 2115, 2116, 2117, 2118, 2119, 2120, 2121, 2122, 2123, 2124, 2125, 2126, 2127, 2128, 2129, 2130, 2131, 2132, 2133, 2134, 2135, 2136, 2137, 2138, 2139, 2140, 2141, 2142, 2143, 2144, 2145, 2146, 2147, 2148, 2149, 2150, 2151, 2152, 2153, 2154, 2155, 2156, 2157, 2158, 2159, 2160, 2161, 2162, 2163, 2164, 2165, 2166, 2167, 2168, 2169, 2170, 2171, 2172, 2173, 2174, 2175, 2176, 2177, 2178, 2179, 2180, 2181, 2182, 2183, 2184, 2185, 2186, 2187, 2188, 2189, 2190, 2191, 2192, 2193, 2194, 2195, 2196, 2197, 2198, 2199, 2200, 2201, 2202, 2203, 2204, 2205, 2206, 2207, 2208, 2209, 2210, 2211, 2212, 2213, 2214, 2215, 2216, 2217, 2218, 2219, 2220, 2221, 2222, 2223, 2224, 2225, 2226, 2227, 2228, 2229, 2230, 2231, 2232, 2233, 2234, 2235, 2236, 2237, 2238, 2239, 2240, 2241, 2242, 2243, 2244, 2245, 2246, 2247, 2248, 2249, 2250, 2251, 2252, 2253, 2254, 2255, 2256, 2257, 2258, 2259, 2260, 2261, 2262, 2263, 2264, 2265, 2266, 2267, 2268, 2269, 2270, 2271, 2272, 2273, 2274, 2275, 2276, 2277, 2278, 2279, 2280, 2281, 2282, 2283, 2284, 2285, 2286, 2287, 2288, 2289, 2290, 2291, 2292, 2293, 2294, 2295, 2296, 2297, 2298, 2299, 2300, 2301, 2302, 2303, 2304, 2305, 2306, 2307, 2308, 2309, 2310, 2311, 2312, 2313, 2314, 2315, 2316, 2317, 2318, 2319, 2320, 2321, 2322, 2323, 2324, 2325, 2326, 2327, 2328, 2329, 2330, 2331, 2332, 2333, 2334, 2335, 2336, 2337, 2338, 2339, 2340, 2341, 2342, 2343, 2344, 2345, 2346, 2347, 2348, 2349, 2350, 2351, 2352, 2353, 2354, 2355, 2356, 2357, 2358, 2359, 2360, 2361, 2362, 2363, 2364, 2365, 2366, 2367, 2368, 2369, 2370, 2371, 2372, 2373, 2374, 2375, 2376, 2377, 2378, 2379, 2380, 2381, 2382, 2383, 2384, 2385, 2386, 2387, 2388, 2389, 2390, 2391, 2392, 2393, 2394, 2395, 2396, 2397, 2398, 2399, 2400, 2401, 2402, 2403, 2404, 2405, 2406, 2407, 2408, 2409, 2410, 2411, 2412, 2413, 2414, 2415, 2416, 2417, 2418, 2419, 2420, 2421, 2422, 2423, 2424, 2425, 2426, 2427, 2428, 2429, 2430, 2431, 2432, 2433, 2434, 2435, 2436, 2437, 2438, 2439, 2440, 2441, 2442, 2443, 2444, 2445, 2446, 2447, 2448, 2449, 2450, 2451, 2452, 2453, 2454, 2455, 2456, 2457, 2458, 2459, 2460, 2461, 2462, 2463, 2464, 2465, 2466, 2467, 2468, 2469, 2470, 2471, 2472, 2473, 2474, 2475, 2476, 2477, 2478, 2479, 2480, 2481, 2482, 2483, 2484, 2485, 2486, 2487, 2488, 2489, 2490, 2491, 2492, 2493, 2494, 2495, 2496, 2497, 2498, 2499, 2500, 2501, 2502, 2503, 2504, 2505, 2506, 2507, 2508, 2509, 2510, 2511, 2512, 2513, 2514, 2515, 2516, 2517, 2518, 2519, 2520, 2521, 2522, 2523, 2524, 2525, 2526, 2527, 2528, 2529, 2530, 2531, 2532, 2533, 2534, 2535, 2536, 2537, 2538, 2539, 2540, 2541, 2542, 2543, 2544, 2545, 2546, 2547, 2548, 2549, 2550, 2551, 2552, 2553, 2554, 2555, 2556, 2557, 2558, 2559, 2560, 2561, 2562, 2563, 2564, 2565, 2566, 2567, 2568, 2569, 2570, 2571, 2572, 2573, 2574, 2575, 2576, 2577, 2578, 2579, 2580, 2581, 2582, 2583, 2584, 2585, 2586, 2587, 2588, 2589, 2590, 2591, 2592, 2593, 2594, 2595, 2596, 2597, 2598, 2599, 2600, 2601, 2602, 2603, 2604, 2605, 2606, 2607, 2608, 2609, 2610, 2611, 2612, 2613, 2614, 2615, 2616, 2617, 2618, 2619, 2620, 2621, 2622, 2623, 2624, 2625, 2626, 2627, 2628, 2629, 2630, 2631, 2632, 2633, 2634, 2635, 2636, 2637, 2638, 2639, 2640, 2641, 2642, 2643, 2644, 2645, 2646, 2647, 2648, 2649, 2650, 2651, 2652, 2653, 2654, 2655, 2656, 2657, 2658, 2659, 2660, 2661, 2662, 2663, 2664, 2665, 2666, 2667, 2668, 2669, 2670, 2671, 2672, 2673, 2674, 2675, 26

Preparation of Amoxicillin from p-HQM and p-APA

1997

1. *Pharmaceutical Innovation and the Role of Government*

1. 2. 3. 4. 5. 6. 7. 8. 9. 10. 11. 12. 13. 14. 15. 16. 17. 18. 19. 20. 21. 22. 23. 24. 25. 26. 27. 28. 29. 30. 31. 32. 33. 34. 35. 36. 37. 38. 39. 40. 41. 42. 43. 44. 45. 46. 47. 48. 49. 50. 51. 52. 53. 54. 55. 56. 57. 58. 59. 60. 61. 62. 63. 64. 65. 66. 67. 68. 69. 70. 71. 72. 73. 74. 75. 76. 77. 78. 79. 80. 81. 82. 83. 84. 85. 86. 87. 88. 89. 90. 91. 92. 93. 94. 95. 96. 97. 98. 99. 100. 101. 102. 103. 104. 105. 106. 107. 108. 109. 110. 111. 112. 113. 114. 115. 116. 117. 118. 119. 120. 121. 122. 123. 124. 125. 126. 127. 128. 129. 130. 131. 132. 133. 134. 135. 136. 137. 138. 139. 140. 141. 142. 143. 144. 145. 146. 147. 148. 149. 150. 151. 152. 153. 154. 155. 156. 157. 158. 159. 160. 161. 162. 163. 164. 165. 166. 167. 168. 169. 170. 171. 172. 173. 174. 175. 176. 177. 178. 179. 180. 181. 182. 183. 184. 185. 186. 187. 188. 189. 190. 191. 192. 193. 194. 195. 196. 197. 198. 199. 200. 201. 202. 203. 204. 205. 206. 207. 208. 209. 210. 211. 212. 213. 214. 215. 216. 217. 218. 219. 220. 221. 222. 223. 224. 225. 226. 227. 228. 229. 230. 231. 232. 233. 234. 235. 236. 237. 238. 239. 240. 241. 242. 243. 244. 245. 246. 247. 248. 249. 250. 251. 252. 253. 254. 255. 256. 257. 258. 259. 260. 261. 262. 263. 264. 265. 266. 267. 268. 269. 270. 271. 272. 273. 274. 275. 276. 277. 278. 279. 280. 281. 282. 283. 284. 285. 286. 287. 288. 289. 290. 291. 292. 293. 294. 295. 296. 297. 298. 299. 300. 301. 302. 303. 304. 305. 306. 307. 308. 309. 310. 311. 312. 313. 314. 315. 316. 317. 318. 319. 320. 321. 322. 323. 324. 325. 326. 327. 328. 329. 330. 331. 332. 333. 334. 335. 336. 337. 338. 339. 340. 341. 342. 343. 344. 345. 346. 347. 348. 349. 350. 351. 352. 353. 354. 355. 356. 357. 358. 359. 360. 361. 362. 363. 364. 365. 366. 367. 368. 369. 370. 371. 372. 373. 374. 375. 376. 377. 378. 379. 380. 381. 382. 383. 384. 385. 386. 387. 388. 389. 390. 391. 392. 393. 394. 395. 396. 397. 398. 399. 400. 401. 402. 403. 404. 405. 406. 407. 408. 409. 410. 411. 412. 413. 414. 415. 416. 417. 418. 419. 420. 421. 422. 423. 424. 425. 426. 427. 428. 429. 430. 431. 432. 433. 434. 435. 436. 437. 438. 439. 440. 441. 442. 443. 444. 445. 446. 447. 448. 449. 450. 451. 452. 453. 454. 455. 456. 457. 458. 459. 460. 461. 462. 463. 464. 465. 466. 467. 468. 469. 470. 471. 472. 473. 474. 475. 476. 477. 478. 479. 480. 481. 482. 483. 484. 485. 486. 487. 488. 489. 490. 491. 492. 493. 494. 495. 496. 497. 498. 499. 500. 501. 502. 503. 504. 505. 506. 507. 508. 509. 510. 511. 512. 513. 514. 515. 516. 517. 518. 519. 520. 521. 522. 523. 524. 525. 526. 527. 528. 529. 530. 531. 532. 533. 534. 535. 536. 537. 538. 539. 540. 541. 542. 543. 544. 545. 546. 547. 548. 549. 550. 551. 552. 553. 554. 555. 556. 557. 558. 559. 560. 561. 562. 563. 564. 565. 566. 567. 568. 569. 570. 571. 572. 573. 574. 575. 576. 577. 578. 579. 580. 581. 582. 583. 584. 585. 586. 587. 588. 589. 590. 591. 592. 593. 594. 595. 596. 597. 598. 599. 600. 601. 602. 603. 604. 605. 606. 607. 608. 609. 610. 611. 612. 613. 614. 615. 616. 617. 618. 619. 620. 621. 622. 623. 624. 625. 626. 627. 628. 629. 630. 631. 632. 633. 634. 635. 636. 637. 638. 639. 640. 641. 642. 643. 644. 645. 646. 647. 648. 649. 650. 651. 652. 653. 654. 655. 656. 657. 658. 659. 660. 661. 662. 663. 664. 665. 666. 667. 668. 669. 670. 671. 672. 673. 674. 675. 676. 677. 678. 679. 680. 681. 682. 683. 684. 685. 686. 687. 688. 689. 690. 691. 692. 693. 694. 695. 696. 697. 698. 699. 700. 701. 702. 703. 704. 705. 706. 707. 708. 709. 710. 711. 712. 713. 714. 715. 716. 717. 718. 719. 720. 721. 722. 723. 724. 725. 726. 727. 728. 729. 730. 731. 732. 733. 734. 735. 736. 737. 738. 739. 740. 741. 742. 743. 744. 745. 746. 747. 748. 749. 750. 751. 752. 753. 754. 755. 756. 757. 758. 759. 760. 761. 762. 763. 764. 765. 766. 767. 768. 769. 770. 771. 772. 773. 774. 775. 776. 777. 778. 779. 780. 781. 782. 783. 784. 785. 786. 787. 788. 789. 790. 791. 792. 793. 794. 795. 796. 797. 798. 799. 800. 801. 802. 803. 804. 805. 806. 807. 808. 809. 810. 811. 812. 813. 814. 815. 816. 817. 818. 819. 820. 821. 822. 823. 824. 825. 826. 827. 828. 829. 830. 831. 832. 833. 834. 835. 836. 837. 838. 839. 840. 84

$\frac{d}{dt} \left(\frac{1}{\rho} \right) = - \frac{1}{\rho^2} \frac{d\rho}{dt}$

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1. *Journal of the American Medical Association*, 283: 2653-2658, 2000.

TEXT:

TABLE 3 _____ substrate crystal cake in
solution added at the end at the end _____
1-HASH 441.3 g 1.1 g 10.4 g 2017 mmol 1 mmol 133 mmol 6-APA 445.1 g 0.1
14.3 g 0.1 mmol 0.1 mmol 12 mmol Amoxicillin. H.sub.2O 135.4 g 0.4
1.141 mmol 0.1 mmol 1-HH 1.1 g 0.1 g 0.1 mmol 177 mmol
Amoxicillin yield is 14.3 g 14.3%,
based on 6-APA added: 17.7%. Amoxicillin productivity: 1.1 mmol/liter. h =
1.4.

TEXT:

TABLE 3 _____ substrate crystal at in
solution added the end at the end _____
1-HASH 441.3 g 1.1 g 10.4 g 2017 mmol 1 mmol 133 mmol 6-APA 445.1 g 0.1
14.3 g 0.1 mmol 0.1 mmol 12 mmol Amoxicillin. H.sub.2O 135.4 g 0.4
1.141 mmol 0.1 mmol 1-HH 1.1 g 0.1 g 0.1 mmol 177 mmol
Amoxicillin yield is 14.3 g 14.3%,
based on 6-APA added: 17.7%. Amoxicillin productivity: 1.1 mmol/liter. h =
1.4.

CLAIM:

14. The process according to claim 1, wherein the amino .beta.-lactam is
selected from the group consisting of 7-aminocapillanic acid,
7-amino-deaacetoxycephaleosporanic acid, 7-aminocephaleosporanic acid,
7-amino-3-chloro-3-cephem-4-carboxylate and 7-amino-3-3-cephem-4-carboxylate.

CLAIM:

15. The process according to claim 1, wherein the acylating agent is selected
from the group consisting of an activated form of D phenylglycine, D
p-hydroxyphenylglycine, D-2,5-dihydroxyphenylglycine and mandelic acid.

CLAIM:

16. The process according claim 1, wherein the .beta.-lactam compound is
selected from the group consisting of ampicillin, amoxicillin, cefaclor,
cephalexin, cephadroxil, cephadrine, epicillin and cefamandel.

WEST**End of Result Set**

Generate Collection

Title: Entry 12 - 12

File: INFO

Rev: 1, 1993

INVENT-ASSN-NR: 12345-1234

INVENT-WHERE: 12345

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TITLE: Recovery of 5-phenyl-glycine amide from antibiotic coupling - by
 formation of pure, easily sepd. recyclable Schiff base, used in asymmetric prepn
 of cephalosin, cefaclor, ampicillin, etc.

INVENTOR: P. BROWN, M. H. T.; M. JY, H. M.; M. JY, H.

INVENT-ASSN-NR:

ASSIGNER

CODE

DSM NR

SIAM

PRIORITY-DATA:

1993BR-1001781

July 19, 1993

INVENT-BASIC:

INVENT-NR

INVENT-DATE

LANGUAGE

PAGES

MAIN-123

ES 2110251 T3

February 1, 1993

N/A

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C12P035/04

WO 9503420 A1

February 2, 1995

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C12P035/04

BE 1007296 A3

May 9, 1995

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C12P000/00

EP 712443 A1

May 22, 1996

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C12P035/04

IN 1111111 A

July 14, 1996

N/A

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C12P035/04

EP 712443 A1

November 14, 1997

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C12P035/04

IN 1111111 A

January 1, 1998

N/A

C12P035/04

DESIGNATED-STATES: CN JP KR US AT BE CH DE DK ES FR GB GR IE IT LU NL PT SP
 AT DE ES FR GB IT NL PT AT DE ES FR GB IT NL PT

ATTACHED-DOCUMENTS: EP 442894; EP 442899; US 4118940; N/A 442894; N/A 442899

INVENT-ASSN-NR:

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|----------------|----------------|----------------|----------|
| EP-0 | AP11-1477111-0 | AP11-0 | AP11-0 |
| EP 711443B | July 18, 1994 | EP 711443B | N/A |
| EP 711443B | N/A | EP 711443B | Based on |
| W. 8013421A1 | July 18, 1994 | 1994WI-NL11161 | N/A |
| EP 117128A3 | July 18, 1993 | 1993BE-1111781 | N/A |
| EP 711443A1 | July 18, 1994 | 1994BE-1924411 | N/A |
| EP 711443A1 | July 18, 1994 | 1994BE-NL11161 | N/A |
| EP 711443A1 | N/A | W. 8013421 | Based on |
| EP 711443A1 | July 18, 1994 | 1994WI-NL11161 | N/A |
| EP 711443B1 | July 18, 1994 | 1994BE-1924411 | N/A |
| EP 711443B1 | July 18, 1994 | 1994WI-NL11161 | N/A |
| EP 711443B1 | N/A | W. 8013421 | Based on |
| 1994WI-NL11161 | July 18, 1994 | 1994BE-1924411 | N/A |
| 1994WI-NL11161 | July 18, 1994 | 1994BE-1924411 | N/A |
| 1994WI-NL11161 | July 18, 1994 | 1994WI-NL11161 | N/A |
| 1994WI-NL11161 | N/A | EP 711443B | Based on |
| 1994WI-NL11161 | N/A | W. 8013421 | Based on |

INT-CL (IPC): G07C 231/20; G12B 1/01; G12B 17/10; G12B 35/04; G12B 37/04

ABSTRACTED-FUB-NO: EP 712443B

BASIC-ABSTRACT:

Prepn of a beta-lactam deriv. is claimed in which a beta-lactam nucleus is coupled to L-phenylglycine amide in an enzymatic reaction, and the enzyme, solid L-phenylglycine, and beta-lactam deriv are sepd out. After the enzymatic reaction, the solid mixt (from which at least the enzyme and solid L-phenylglycine have been removed), is treated with an aldehyde at a pH of 7.8-8.8 and the Schiff base of L-phenylglycine amide is sepd. out.

USE - The coupling reaction is used for prepn of important antibiotics, including ampicillin, cephalixin and cephalexin, in which the beta-lactam nucleus are respectively penicillanic acid, 6-APA, 7-aminocaproxam, 7-aminocaproxam, and 7-aminocaproxam, and 7-aminocaproxam is coupled with L-phenylglycine, and related acids.

ADVANTAGE - The L-phenylglycine amide deriv is easily sepd. out in pure form, pref before the beta-lactam prod as the solubility of the latter is higher; recovery and purification of this prod by known methods, is simplified. Recovery and recycle of the L-phenylglycine amide deriv is by simple filtration or evapn. In particular if the aldehyde benzaldehyde is used, excess aldehyde can serve as extn solvent, although other solvents, or mixts with EtOH, can be used. The Schiff base is then split with acid, eg HCl, and recycled. As an excess of amine is used in the coupling to obtain a high yield of beta-lactam prod, recovery of the amine is necessary in order to provide a commercially attractive process; the reaction mixt typically contains 1-7 moles of L-phenylglycine amide, 1.1-1.5 moles of L-phenylglycine and 1.1-1.5 equiv of activated lactam, all per mole of aldehyde.

ABSTRACTED-FUB-NO:

W. 8013421A

BASIC-ABSTRACT:

Prepn of a beta-lactam deriv. is claimed in which a beta-lactam nucleus is coupled to L-phenylglycine amide in an enzymatic reaction, and the enzyme, solid L-phenylglycine, and beta-lactam deriv are sepd out. After the enzymatic reaction, the solid mixt (from which at least the enzyme and solid L-phenylglycine have been removed), is treated with an aldehyde at a pH of

1 of 1

UNLINKED-DERWENT-REGISTRY-NUMBERS: 0289P: 1986P

SET NEARY-APP-N :

THE Secondary Addressed to Name-ys: 71000-198600

WEST

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NO.: Entry 4 of 11

File: WEST

Nov 10, 1977

SUBJECT-IDENTIFIER: 12-000000 A

TITLE: Penicillin and cephalosporin intermediates

ABST:

Novel penicillin and cephalosporin intermediates for the synthesis of α -APA or β -aminocephem compounds by living reaction of α -APA or β -aminocephem phosphite acid compound and the conversion of such compound to a desired α -APA or β -aminocephem compound.

BRIEF:

Numerous methods for the synthesis of penicillins or cephalosporins have been described in the literature. The major part of these syntheses involves the reaction of β -amino penicillanic acid (in the following referred to as β -APA), salts and esters thereof or the corresponding β -aminocephem compounds with an activated derivative of the acid, with which the amino group is to be acylated.

BSPR:

In one aspect of the invention there is provided a novel intermediate for the synthesis of derivatives of α -APA or β -aminocephem compounds in high yields.

BSPR:

The structure of the compound having formula I has been established in the case of both ^{13}C and ^{31}P spectra (see Example 1). The spin-spin coupling pattern of the latter shows that the α -alpha-hydrogen of the α -APA derivative or the corresponding β -alpha-hydrogen of the β -aminocephem derivative is strongly influenced by the introduction of the phosphorus atom.

BSPR:

In α -APA derivatives with a free NH-substituent-group the proton in the α -position is usually seen as a doublet at δ 6.1-6.4 - 6.5 ppm (CDCl₃ soln.). However, as an example in α -APA compounds having the formula I, cf. Example 1, this proton is found as a multiplet consisting of 4 peaks in the region of δ 6.1-6.5 - 6.6 ppm (CDCl₃ soln.), from which by first order analysis the following spin-spin coupling constants can be obtained:

BSPR:

It is even more surprising that a compound having the formula I, wherein R is a secondary ammonium group can be reacted with a halophosphite compound to form the corresponding phosphiteamide compound. However, it has been found that for example β -amino penicillanic acid or β -amino desacetoxyccephalosporanic acid, when dissolved in acetonitrile or dimethylformamide in the presence of two molar equivalents of triethylamine reacts with triethylamine chlorophosphite salts to form a compound of the triethylammonium salt of β -triethylphosphite penicillanic acid or β -triethylphosphite amides desacetoxyccephalosporanic acid and diethylammonium chloride. It might have been expected that triethylamine chlorophosphite would react exclusively with the expressive mole of secondary amine.

BSPR:

An alternative procedure in the preparation of penicillin and cephalosporin compounds from β -amino penicillanic acid, β -aminocephem and a phosphite compound, as described in Example 1 is frequently used in the presence of a

1. *Journal of the American Medical Association*, 1997; 277: 1001-1005.

| Year | Age | Sex | Location | Notes |
|------|-----|--------|----------|-------|
| 1998 | 1 | Male | 1000 | 1000 |
| 1999 | 2 | Female | 1000 | 1000 |
| 2000 | 3 | Male | 1000 | 1000 |
| 2001 | 4 | Female | 1000 | 1000 |
| 2002 | 5 | Male | 1000 | 1000 |
| 2003 | 6 | Female | 1000 | 1000 |
| 2004 | 7 | Male | 1000 | 1000 |
| 2005 | 8 | Female | 1000 | 1000 |
| 2006 | 9 | Male | 1000 | 1000 |
| 2007 | 10 | Female | 1000 | 1000 |
| 2008 | 11 | Male | 1000 | 1000 |
| 2009 | 12 | Female | 1000 | 1000 |
| 2010 | 13 | Male | 1000 | 1000 |
| 2011 | 14 | Female | 1000 | 1000 |
| 2012 | 15 | Male | 1000 | 1000 |
| 2013 | 16 | Female | 1000 | 1000 |
| 2014 | 17 | Male | 1000 | 1000 |
| 2015 | 18 | Female | 1000 | 1000 |
| 2016 | 19 | Male | 1000 | 1000 |
| 2017 | 20 | Female | 1000 | 1000 |
| 2018 | 21 | Male | 1000 | 1000 |
| 2019 | 22 | Female | 1000 | 1000 |
| 2020 | 23 | Male | 1000 | 1000 |
| 2021 | 24 | Female | 1000 | 1000 |
| 2022 | 25 | Male | 1000 | 1000 |
| 2023 | 26 | Female | 1000 | 1000 |
| 2024 | 27 | Male | 1000 | 1000 |
| 2025 | 28 | Female | 1000 | 1000 |
| 2026 | 29 | Male | 1000 | 1000 |
| 2027 | 30 | Female | 1000 | 1000 |
| 2028 | 31 | Male | 1000 | 1000 |
| 2029 | 32 | Female | 1000 | 1000 |
| 2030 | 33 | Male | 1000 | 1000 |
| 2031 | 34 | Female | 1000 | 1000 |
| 2032 | 35 | Male | 1000 | 1000 |
| 2033 | 36 | Female | 1000 | 1000 |
| 2034 | 37 | Male | 1000 | 1000 |
| 2035 | 38 | Female | 1000 | 1000 |
| 2036 | 39 | Male | 1000 | 1000 |
| 2037 | 40 | Female | 1000 | 1000 |
| 2038 | 41 | Male | 1000 | 1000 |
| 2039 | 42 | Female | 1000 | 1000 |
| 2040 | 43 | Male | 1000 | 1000 |
| 2041 | 44 | Female | 1000 | 1000 |
| 2042 | 45 | Male | 1000 | 1000 |
| 2043 | 46 | Female | 1000 | 1000 |
| 2044 | 47 | Male | 1000 | 1000 |
| 2045 | 48 | Female | 1000 | 1000 |
| 2046 | 49 | Male | 1000 | 1000 |
| 2047 | 50 | Female | 1000 | 1000 |
| 2048 | 51 | Male | 1000 | 1000 |
| 2049 | 52 | Female | 1000 | 1000 |
| 2050 | 53 | Male | 1000 | 1000 |
| 2051 | 54 | Female | 1000 | 1000 |
| 2052 | 55 | Male | 1000 | 1000 |
| 2053 | 56 | Female | 1000 | 1000 |
| 2054 | 57 | Male | 1000 | 1000 |
| 2055 | 58 | Female | 1000 | 1000 |
| 2056 | 59 | Male | 1000 | 1000 |
| 2057 | 60 | Female | 1000 | 1000 |
| 2058 | 61 | Male | 1000 | 1000 |
| 2059 | 62 | Female | 1000 | 1000 |
| 2060 | 63 | Male | 1000 | 1000 |
| 2061 | 64 | Female | 1000 | 1000 |
| 2062 | 65 | Male | 1000 | 1000 |
| 2063 | 66 | Female | 1000 | 1000 |
| 2064 | 67 | Male | 1000 | 1000 |
| 2065 | 68 | Female | 1000 | 1000 |
| 2066 | 69 | Male | 1000 | 1000 |
| 2067 | 70 | Female | 1000 | 1000 |
| 2068 | 71 | Male | 1000 | 1000 |
| 2069 | 72 | Female | 1000 | 1000 |
| 2070 | 73 | Male | 1000 | 1000 |
| 2071 | 74 | Female | 1000 | 1000 |
| 2072 | 75 | Male | 1000 | 1000 |
| 2073 | 76 | Female | 1000 | 1000 |
| 2074 | 77 | Male | 1000 | 1000 |
| 2075 | 78 | Female | 1000 | 100 |

$\frac{1}{2} \times \frac{1}{2} = \frac{1}{4}$

[illegible]

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$\frac{1}{2} \times \frac{1}{2} = \frac{1}{4}$

Figure 1

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stirred for 10 minutes at room temperature and filtered and washed.

ANAL:

dry ether is added to the mixture to precipitate the triethylammonium chloride salt. The salt is filtered off. The filtrate is evaporated to a white wax which is very hygroscopic. The product is stable in cold, dry solution and when stored under dry nitrogen. The product may contain small amounts of unreacted p-aminophenylalanine acid, triethylammonium salt.

ANAL:

1.0 g (5 millimoles) of p-aminophenylalanine acid are dissolved in 5 ml dry acetonitrile by the addition of 1.4 ml (5 millimoles) of triethylamine. The solution is cooled to -40.degree. C. and while stirring 1.0 ml (5 millimoles) of 4-oxo-1,2,3,4-tetrahydro-1,4-benzoxaphosphinane dissolved in 2 ml anisole are added under dry nitrogen. Stirring is continued for 10 minutes at -40.degree. C. and subsequently 10 minutes at room temperature. 15 ml dry benzene are added. The triethylammonium chloride formed is separated by filtration. During said filtration part of the product is lost, but the filtration is normally unnecessary when the product is to be used for further reactions.

ANAL:

1.0 g (5 millimoles) of p-aminophenylalanine acid are dissolved in 5 ml dry acetonitrile by the addition of 1.4 ml (5 millimoles) of triethylamine while stirring. 1.4 ml (5 millimoles) of 8-chloro-1,3,2-thioxaphospholan in 2 ml dry acetonitrile are added dropwise under dry nitrogen and at a temperature of about -40.degree. C. Subsequently the solution is stirred for about 1 hour at room temperature.

ANAL:

1.03 ml (10 millimoles) of diethylamine are added to 1.03 g (5 millimoles) of p-aminophenylalanine acid suspended in 10 ml dry acetonitrile. After stirring in a nitrogen atmosphere for 2 minutes, a clear solution is obtained.

ANAL:

To a suspension of 1.0 g (5 millimoles) of p-APA in 10 ml of dry methylene chloride are added 1.4 ml (5 millimoles) of triethylamine, and the mixture is stirred at room temperature until a clear solution is formed. The solution is cooled to -40.degree. C. and 1.48 ml (5 millimoles) of ethylenedichlorophosphite dissolved in 5 ml of dry methylene chloride is added under dry nitrogen. The mixture is stirred for 10 minutes at -40.degree. C., 15 minutes at 0.degree. C. and 10 minutes at room temperature, thereafter 2.67 ml (5.3 millimoles) of trimethylchlorosilane dissolved in 5 ml of methylene chloride is added. The mixture is stirred for 5 minutes at room temperature, and subsequently the precipitated triethylamine hydrochloride is filtered off. The solution is evaporated to form a hard, pale yellow oil.

ANAL:

1 g of p-APA is suspended in 10 ml of dry methylene chloride under nitrogen. To this mixture are added 1.48 ml of dry triethylamine and 1.61 ml of trimethylchlorosilane. After stirring for 1 hour the reaction mixture is cooled to -40.degree. C. and a solution of 2.67 ml of p-phenylenedichlorophosphite in 5 ml of dry methylene chloride is added dropwise. When the addition is complete the stirring is stopped, the mixture is stirred for 4 minutes, and the solvent is removed in vacuo. Dry benzene is added, and the triethylammonium chloride formed is removed by filtration under nitrogen. The benzene is removed in vacuo. The residue is a solid which is purified by recrystallization as an amorphous substance.

ANAL:

10.71 g (50 millimoles) of D-(-)-beta-phenylalanylchloride, HCl are then added to the reaction mixture, and the mixture is stirred for 1 hour at -40.degree. C. Subsequently 10 ml water is added and the mixture is stirred for 10 minutes at room temperature. The pH is adjusted to 7.0 with 10% sodium hydroxide solution and the reaction mixture is filtered

the 1st phase which is then slowly washed with water. The organic phase is separated and the aqueous phase is washed twice with 10 ml methylene chloride. The organic phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 2nd phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 3rd phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 4th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 5th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 6th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 7th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 8th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 9th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 10th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 11th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 12th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 13th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 14th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 15th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 16th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 17th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 18th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 19th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 20th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 21st phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 22nd phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 23rd phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 24th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 25th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 26th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 27th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 28th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 29th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 30th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 31st phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 32nd phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 33rd phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 34th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. 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The 47th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 48th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 49th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 50th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 51st phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 52nd phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 53rd phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 54th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 55th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 56th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 57th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 58th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 59th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 60th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 61st phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 62nd phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 63rd phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 64th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 65th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 66th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 67th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 68th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 69th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 70th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 71st phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 72nd phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 73rd phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 74th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 75th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 76th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 77th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 78th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 79th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 80th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 81st phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 82nd phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 83rd phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 84th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 85th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 86th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 87th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 88th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 89th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 90th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 91st phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 92nd phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 93rd phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 94th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 95th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 96th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 97th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 98th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 99th phase is washed with 10 ml water and dried with anhydrous sodium sulfate. The 100th phase is washed with 10 ml water and dried with anhydrous sodium sulfate.

BSFR:

4.33 g (21 millimoles) of D(-)-phenylglycylchloride, hydrochloride are added in portions over a period of 1 minute to a solution in methylene chloride of 21 millimoles of trimethylsilyl-6-ethylenephosphiteamidepenicillanate prepared as described in Example 7 and without leaving the temperature at 0 degree. C. The progress of the reaction is followed by penicillinase titration which after a period of 2 hours at 0 degree. C shows that a yield of above 70% has been obtained. At this time the reaction mixture is poured into 100 ml ice-water, and after stirring for 15 minutes the reaction mixture is filtered. The water phase is covered by 20 ml ethylacetate and the pH-value of the water phase is adjusted at 2 with a NaOH solution. Subsequently, the ampicillin formed is precipitated as a sparingly soluble salt with .beta.-naphthalenesulphonic acid while maintaining the pH-value at 2. The reaction mixture is left to stand for 17 hours at 0 degree. C and is subsequently filtered and the residue is washed with 10 ml 0.1N HCl and ethylacetate. After drying in vacuum 7.2 g white ampicillin .beta.-naphthalenesulphonic acid salt corresponding to 64.5% of the theoretical amount are obtained. A high-voltage electrophogram showed one spot with mobility identical to that of authentic ampicillin. The IR spectrum was identical to that of the .beta.-naphthalenesulphonic acid salt of authentic ampicillin.

BSFR:

1.0 g (5 millimoles) of 1-methoxy-2-methyl-2-propyl-3-oxo-1,2,3,4-tetrahydropyridine-4-carboxylic acid are dissolved in 10 ml of dry methylene chloride and heated to reflux. A solution of 1.0 g (5 millimoles) of trimethylsilyl-6-ethylenephosphiteamidepenicillanate in 10 ml of dry methylene chloride is added during 15 minutes. Progress titration 15 minutes after the addition is completed shows a penicillin yield of 80%, while titration after one and two hours both shows 80% yield.

BSFR:

0.33 g (1.65 millimoles) of 1-methoxy-2-methyl-2-propyl-3-oxo-1,2,3,4-tetrahydropyridine-4-carboxylic acid are dissolved in 10 ml of dry methylene chloride and heated to reflux. A solution of 0.33 g (1.65 millimoles) of trimethylsilyl-6-ethylenephosphiteamidepenicillanate in 10 ml of dry methylene chloride is added during 15 minutes. Progress titration 15 minutes after the addition is completed shows a penicillin yield of 80%, while titration after one and two hours both shows 80% yield.

BSFR:

4 millimoles of N-tert.-butoxycarbonyl-D(-)-.alpha.-phenylglycine trimethylsilyl-ester prepared from 1.1 g (4 millimoles) of N-tert.-butoxycarbonyl-D(-)-.alpha.-phenylglycine, 10 ml of chloroform, 0.1 g (4 millimoles) of triethylamine and 0.1 g (4 millimoles) of trimethylsilyl-6-ethylenephosphiteamide are added to a solution of 4

1. The first step in the process of preparing a solution is to determine the concentration of the solution. This is done by measuring the mass of the solute and the volume of the solvent. The concentration is then expressed as a percentage or a molar concentration.

2. The second step is to weigh the solute. This is done using a balance scale. The mass of the solute is then divided by the volume of the solvent to give the concentration.

3. The third step is to add the solute to the solvent. This is done by pouring the solvent into a beaker and then adding the solute. The mixture is then stirred to ensure that the solute is fully dissolved.

4. The fourth step is to transfer the solution to a volumetric flask. This is done by pouring the solution from the beaker into the flask. The flask is then filled with solvent to the mark on the neck of the flask.

5. The fifth step is to stopper the flask and invert it several times to ensure that the solution is well mixed.

6. The sixth step is to label the flask with the concentration of the solution.

7. The seventh step is to store the solution in a cool, dark place.

8. The eighth step is to use the solution as required.

9. The ninth step is to dispose of the solution properly.

10. The tenth step is to clean the glassware used in the process.

To a solution of 10 millimoles of 2-oxo-2-phenyl-3-(4-methylphenyl)-5-oxaphosphorin-3-ylidene-1-penicillinate prepared as described in Example 1, 100 ml. of *n*-BuOH, 100 ml. of dry methylene chloride are added, 0.1 g. of 10 millimoles of phenylphosphonic acid, and the mixture is stirred at room temperature with dry air bubbling through. After a reaction period of 2 hours at room temperature the reaction mixture shows a penicillin yield of 50% by gravimetric titration. The reaction mixture is cooled to 0 degree C, and 5 ml. of pyridine are added followed by 25 ml. of dimethylformamide, whereafter the reaction mixture is poured into 500 ml. of ice-cold 1% NaCl solution and stirred for 30 minutes. Then 150 ml. of ethylacetate are added and the pH adjusted to 2. After 30 minutes the phases are separated, and the water phase is extracted three times with ethylacetate. 25 ml. of water are added to the combined organic phases, and the pH is adjusted to 7 with KOH. The water phase is separated, 150 ml. of *n*-butanol added and the water removed by azeotropic vacuum distillation. The crystalline crude product thus precipitated is filtered off. Yield: 3.16 g. (81%) of a purity of 76% determined by penicillinase titration. The white, crystalline product shows a characteristic absorption in the IR spectrum corresponding to that of the potassium salt of phenylmethylpenicillin, and the NMR spectrum also shows signals which are characteristic of said compound.

1.4 g (4 millimoles) of phenylacetic acid is added to a solution of 1,1,1-trichloroethyl-3-methyl-7-oxa-10-phenylphosphorothioate-*n*-4-carboxylate in acetanilide prepared as described in Example 11, except for the addition. Remove the precipitate of triethylammonium chloride. The mixture is stirred for 16 hours at room temperature. The reaction mixture is then poured into 100-ml water and extracted three times with methyl acetate. The organic phases are washed three times with 10% dilute sodium bicarbonate solution and three times with saturated sodium hydroxide carbonate solution, dried over magnesium sulfate and evaporated in vacuum to form an amorphous powder:

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1.1 g (4 millimoles) of N-(tert-butoxycarbonyl)-D,L'-alpha.-isopropyl-L-proline are added to a solution of 0.7 g (1.1 millimoles) of triethylamine in 10 ml of dichloromethane. 1.0 g (4 millimoles) of 1,1'-bis(hydroxyethyl)-3-propanol-7-butyne-1-ethylethylphosphonamide-1-eph-1-ol are added to the mixture, prepared as described in Example 1, except for the addition of one of the previously mentioned phosphonate diluents. The mixture is stirred for 24 hours. Then the reaction mixture is poured into 100 ml of water and extracted three times with diethyl ether. The combined organic

phases are washed three times with 10 ml of distilled water and three times with 10 ml of 10% aqueous sodium bicarbonate solution, dried over magnesium sulfate and concentrated under reduced pressure to give 0.5 g of 6-APA.

BSPP:

1.0 g of 6-aminopenicillanic acid are dissolved in 25 ml of acetonitrile by addition of 10 ml of triethylamine, and the solution formed is cooled to -10 degrees C. A solution of 4.4 ml of ethylenediphosphite in 25 ml of acetonitrile cooled to -10 degrees C is added in the portion while stirring and the stirring is continued for 1 hour. During which period, the temperature gradually rises to 0 degrees C. The triethylamine salt is removed by filtration.

BSPP:

A solution of the triethylammonium salt of 6-ethylene phosphite amidopenicillanic acid was prepared from 4.16 g (15 millimoles) of 6-aminopenicillanic acid in the manner described in Example 1 and the triethylammonium chloride formed was removed by filtration.

BSPP:

The NMR spectrum (H.sub.3 CN) of a sample of the reaction solution showed that all 6-aminopenicillanic acid had been converted into the corresponding 6-ethylene phosphite amide compound, and the following characteristic signals were obtained:

BSPL:

has the meaning defined above with a salt of 6-APA or of 7-aminocephem acid or a derivative of said acids.

BSPL:

The coupling constants J.sub.P-N-C-H, J.sub.H-N-C-H and J.sub.H-C-C-H are consistent with those found in literature (for P-N-CH.sub.3 compounds are normally found J.sub.P-N = 4.5 - 25 Hz (Jackman and Sternhell: Applications of Nuclear Magnetic Resonance Spectroscopy in Organic Chemistry, vol. 5, p. 351 (1969)). The coupling J.sub.H-C-C-H = 4.5 Hz is also found in the signal "doublet" from the proton in the 6-position in 6-APA. This pattern in conjunction with the signal of the 3-hydrogen and the infrared absorption frequencies of the carbonyl group in the 6-position which are consistent with those normally found in 6-APA derivatives, is a conclusive proof for the presence of the structural element: **STR6**

BSPP:

6-APA (1.0 g)

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Doc Entry: 100000000

Entry Title:

May 2, 1981

SUBJECT-IDENTIFIER: US 441-55-A

TITLE: Process for preparing cephalosporanic acid compounds

RIIR:

It is known from the literature to prepare penicillanic acid and cephalosporanic acid derivatives by acylating α -AAA or β -AAA and their derivatives with the hydrochloride of α -benzylglycine-chloride and derivatives thereof having a substituted phenyl group, whereby the said acid chloride is obtained by reacting the substituted phenylglycine with reagents like phosphorus pentachloride, thionyl chloride and phosgene. Although improved processes for the preparation of D-(-)-2-(p-hydroxyphenyl)-glycyl-chloride hydrochloride and the crystalline hemidioxane solvate thereof are known from British Pat. No. 1,466,637 and No. 1,460,915 the acylation of α -AAA or β -AAA or its β -methyl modification with the above-mentioned acylating agent did not hitherto lead to results aimed at, mainly because either the product formed was too impure that further recovery of a product of the required quality hardly appeared to be possible, or the starting D-(-)-2-(p-hydroxyphenyl)-glycyl-chloride hydrochloride of the required quality (purity) is only available for economically unattractive prices, if at all.

RSPR:

A process for the preparation of amoxicillintrihydrate is also known from published German patent application DT 261286 comprising the acylation of α -AAA, which has been previously silylated, with D-(-)-2-(p-hydroxyphenyl)-glycyl-chloride hydrochloride and leading to yields, which seem to approach to some extent the present practical requirements.

RSPR:

However, in the preparation of D-(-)-2-(p-hydroxyphenyl)-glycyl-chloride hydrochloride according to the British Pat. No. 1,460,915 and British Pat. No. 1,466,637, phosgene is used in a relatively difficultly manageable process in which a solid is reacted with a gas. Such a process is extremely expensive in a number of countries with very stringent safety regulations, if indeed it may be applied at all. For the same reason, the process described in British Pat. Nos. 1,460,915 and 1,466,637 disclosing the preparation of α -isocyanatopenicillanic acid and β -isocyanatoccephalosporanic acid derivatives from esters of α -AAA or β -AAA or its β -methyl modifications with phosgene and the subsequent reaction to form penicillins or cephalosporins are left out of consideration for the preparation of amoxicillin and its cephalosporanic acid derivatives because the necessary amount of phosgene is required.

RSPR:

Less attractive features of this process are that the process is carried out at low concentrations, that solvents become mixed so that recovery thereof becomes more difficult, and that, when adding the dimethylacetamide hydrochloride, high concentrations, α -AAA, β -AAA or β -AAA derivatives are required, so that a very large amount of waste is required.

RSPR:

Furthermore, a number of patent applications and patents disclose preparation methods of α -aminoacyl-penicillanic acid derivatives by acylating α -AAA with mixed anhydrides derived from modified Dene salts of D-2-amino- p-hydroxyphenyl-acetic acid, such as those described in German

parent Application No. Ser. No. 1,367,342, No. 1,382,285, No. 1,382,286 and No. 1,382,287, and British Pat. Nos. 1,367,342 and No. 1,382,285. However, the yields disclosed in the above-mentioned applications are unsatisfactory as well as the purpose of the present invention, and moreover, the large scale apparatus available in economically attractive quantities, if at all available.

DEPR:

British patent Application No. Ser. No. 1,367,342 further describes the protection of the amino group of 6-APA, 7-APA and other amino acids by reaction of with organosilane derivatives and lower α -functional silicon compounds and reveals apparently that the most satisfactory triorganosilane compounds and the application thereof should, in a number of cases, lead to improved yields, as appears, even as from British Pat. No. 1,367,342 disclosing the preparation of intermediate organosilane penicillins by reaction of 6-APA and those α -functional silicon compounds. The organosilane derivatives are acylated into amoxicillin, for example, so that an expert from the contents of this patent would expect that the use of the organosilane penicillins described therein would lead to interesting yields in the preparation of amoxicillin. However, this expectation could surprisingly not be confirmed by initial experiments.

ESPR:

Although it is further known from a number of patent applications such as Japanese Patent application No. 49-114687 and No. 49-046892, British patents No. 1,367,342 and No. 1,382,285 and German patent applications Ser. No. 2,460,649 and No. 2,621,618, to prepare amoxicillin from 6-APA and 6-hydroxy-phenylglycine or lower alkyl esters thereof by enzymatic acylation, the processes of this type are also unsatisfactory for the deemed purpose in view of the yields obtained and/or the presence of the acylating enzyme in the amoxicillin-containing solution obtained.

BSFR:

It has been found that the way in which the silylation is carried out is very important for the final yield, and the silylation is preferably carried out in dry methylene chloride containing 2 to 3 equivalents of a tertiary amine such as triethylamine and an equivalent amount of TMCS (about 2 equivalents for amoxicillin and cefadroxil and 3 equivalents for cefatrizine), in such a way that the signal recorded by a pH electrode is adjusted at the end of the reaction at a value of, for example, a pH scale value between 6.8 and 7.2, preferably 6.8 and 7.2, of a Radiometer pH meter type TTT2/C and a Radiometer K14 pH electrode or an Ingold, so-called cold electrode, at a temperature between 15.degree. and 16.degree. C. Therefore, disilylation of e.g. of 6-APA or 7-APA is preferably carried out with practically balanced mutual amounts of tri(lower alkyl) halosilane, such as TMCS, and tertiary amine, such as TEA.

DEPR:

According to a further preferred process, the solution of the amine as prepared is cooled to a temperature of -15.degree. C. or lower and a cooled solution of silylated 6-APA, 7-APA or α -methyl derivatives thereof are added rapidly with stirring as well as possible so that a temperature of -15.degree. C. or -16.degree. C. is reached, whereafter the reaction mixture is stirred for a further 2.5 to 3 hours. An excess of the formula V compound is preferably employed, the excess being dependent on the nature of the substituent of the α -methyl group of the cephalosporanic nucleus. For the preparation of amoxicillin and cefadroxil a small excess is sufficient. In case of cefatrizine, having an acylatable amino group in the heterocyclic ring, at least 2 moles of the compound of formula V will be necessary.

BSFR:

It will be appreciated that some of the most important advantages of the present process is the convenient, reproducible and selective silylation of the amino group of the reactants, and the reaction is being carried out in a concentrated solution of the reactants; given the size of the equipment this will favorably influence the output in kilos per batch; the use of a

| | | | | |
|----|----|----|----|----|
| 1 | 2 | 3 | 4 | 5 |
| 6 | 7 | 8 | 9 | 10 |
| 11 | 12 | 13 | 14 | 15 |

1000

1000

100

1000

10. 11. 2001

1.0 g of 1- α -[1-carboxmethoxypropen-2-yl]-amino-p-hydroxyphenylacetate and 7.0 g of triethylamine in 100 ml of dry methylene chloride. The pH value measured with a Radiometer pH meter type TTT-20 and a Radiometer GK-2411C electrode was adjusted at 6.7 at the end of the silylation reaction.

DEPR:

In the same manner as described in Example 1, amoxicillin trihydrate was obtained in a yield of 81.7% having a purity of 99.5% according to a hydroxylamine method measurement, a biologically measured purity of 97.7%, starting from 40.0 g of potassium.

1.0 g of 1- α -[1-carboxmethoxypropen-2-yl]-amino-p-hydroxyphenylacetate in 100 ml of methylenechloride and 60 ml of triethylamine, 1.0 g of N-methylmorpholine, 40 ml of ethyl chloroformate in 50 ml of methyl isobutylketone, 40 ml of 1-APA in 100 ml of dry methylene chloride, 6.0 g of triethylamine and 52 mmol of trimethylchlorosilane.

DEPR:

1.0 g of 1- α -[1-carboxmethoxypropen-2-yl]-amino-p-hydroxyphenylacetate were weighed into a 1 l reaction vessel and 400 ml of methylene chloride were added thereto. After addition of 20 g of bis(trimethylsilyl)-urea, the mixture was refluxed for about 2.5 hours and the mixture was then cooled to 20.degree. C. The "pH" reading, on the scale of a Radiometer pH meter type TTT 20, connected with a Radiometer GK-2411C electrode, was 6.3.

DEPR:

In the same manner as described in Example 13, 44.3 g of amoxicillin trihydrate having a purity of 97.7% were obtained by reaction of methoxytriethylamine, 1- α -[1-carboxmethoxypropen-2-yl]-amino-p-hydroxyphenylacetate and 35 g of 1-aminopenicillanic acid, previously silylated with 72.5 g of bis(trimethylsilyl)-acetamide instead of the bis(trimethylsilyl)-urea.

DEPR:

8. Silylation of 1-APA

DEPR:

1.0 g of 1-APA was added to 400 ml of methylene chloride. After addition of 40 ml of triethylamine at ambient temperature and under stirring, 50 ml of trimethylchlorosilane are added in about 10 minutes at a temperature of 10.degree.-25.degree. C. After additional stirring for 1 hour the "pH" value is adjusted to a final value of 6.7 by the addition of 4.9 ml of trimethylchlorosilane. The mixture is cooled to -40.degree. C.

DEPR:

In the same manner as described in Examples 13-15, 55.3 g of amoxicillin trihydrate are obtained in a yield of 81.5% having a purity of 99.5% according to a hydroxylamine method measurement, a biologically measured purity of 97.7%, a chemically measured purity of 99.5%, and an optical rotation $[\alpha]_D^{25}$ of +30, starting from 55 g of potassium. 1- α -[1-carboxmethoxypropen-2-yl]-amino-p-hydroxyphenylacetate in 100 ml of methylenechloride and 60 ml of N-methylmorpholine, 1.0 g of N-methylmorpholine, 40 ml of ethyl chloroformate, 35 g of 1-APA in 100 ml of methylenechloride, 40 ml of triethylamine and 35 ml of trimethylchlorosilane. The "pH" value measured with a Radiometer pH meter TTT-20, and a Radiometer GK-2411C electrode was adjusted at 6.7 at the end of the silylation reaction, while the solution of the mixed anhydride as well as the solution of silylated 1-APA were both previously cooled to -40.degree. C. and reacted at -30 to -40.degree. C. for 2 hours. A reaction yield of 81.5%.

DEPR:

In the same manner as described in Examples 1-15, 44.3 g of amoxicillin trihydrate are obtained in a yield of 81.7% having a purity of 99.7% according to a hydroxylamine method measurement, a biologically measured purity of 97.7%, a chemically measured purity of 99.7% and an optical rotation $[\alpha]_D^{25}$ of +30, starting from 55 g of potassium.

1- α , β -bis-(2-hydroxypropen-2-yl)-amino- β -hydroxyphenylacetate in 400 ml of methylene-chloride, 2 ml of N,N-dimethylformamide, 1.5 ml of N-methylmorpholine, 1.5 ml of methylchloroformate, 15 g of δ -ASA in 100 ml of methylene-chloride, 40 ml of triethylamine and 15 ml of triethylchlorosilane. The "pH" value measured with a Radiometer pH meter TTT20, and a Radiometer GK 2412 electrode was adjusted at 6.7 at the end of the silylation reaction, while the solutions of the mixed anhydride and of silylated δ -ASA were pre-cooled to -40.degree. C. and reacted at -40.degree. C. for 2 hours.

DEPU:

In the same manner as described in examples 13-15, amoxicillin was prepared in a 40% acylation yield starting from 55.1 g of potassium

1- α , β -bis-(2-hydroxypropen-2-yl)-amino- β -hydroxyphenylacetate in 400 ml of methylene-chloride, 2 ml of N,N-dimethylformamide, 1.5 ml of N-methylmorpholine, 1.5 ml of methylchloroformate, 15 g of δ -ASA in 100 ml of methylene-chloride, 40 ml of triethylamine and 15 ml of triethylchlorosilane. The "pH" value measured with the same equipment as in the preceding example was adjusted at 6.7 at the end of the silylation reaction, while solutions of the mixed anhydride and of silylated δ -ASA were pre-cooled to -40.degree. C. and reacted at -40.degree. C. for 2 hours.

DEPU:

STEP B: Silylation of δ -ASA

DEPU:

STEP B: Silylation of δ -ASA

DEPU:

STEP B: Silylation of δ -ASA

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These two sets of relations are preserved by the two corresponding mappings, i.e., under the correspondence α and β , the same relation ρ which is a member of \mathcal{R} is preserved. Theorem 1.1 states that, if ρ is a relation in \mathcal{R} , then ρ is preserved by α and β if and only if ρ is an inclusion, equality, extension, combination or intersection property. Another example illustrating the use of the algorithm of Theorem 1.1 is the decomposition of the relation ρ in Figure 1. The group α is defined by the mapping from ρ to ρ' in Figure 2. The group β is defined by the mapping from ρ to ρ'' in Figure 3.

1944:
The following substitutions are used in the Examples: IMP, dimethylformamide,
DMAc, N-methyl pyrrolidone, DMF, N,N-dimethylacetamide,
N-methyl-2-pyrrolidone, THF, tetrahydrofuran,
N-methyl-2-pyrrolidone, DMF, N-methyl-2-pyrrolidone,
N-methyl-2-pyrrolidone, DMF, N-methyl-2-pyrrolidone,
N-methyl-2-pyrrolidone, DMF, N-methyl-2-pyrrolidone,
N-methyl-2-pyrrolidone, DMF, N-methyl-2-pyrrolidone.

epinephrine.

1888:
L-AAA triethylamine salt 3.17 g. dissolved in 40 ml. of dichloromethane is added dropwise at -10 degree. To a solution of ethyl-2-D-1-ethoxycarbonyl-1-methylvinylamino-2-p-ethoxycarbonyloxyphenyl benzoate, prepared in situ from 2.75 g. of L-1-ethoxycarbonyl-1-methylvinylamine-2-p-ethoxycarbonylphenylacetic acid and 1.1 g. of ethyl benzoate in the presence of 5 ml. of triethylamine, in 10 ml. of ethyl acetate. The reaction mixture is allowed to warm slowly to ambient temperature and extracted with 5 ml. of water. The aqueous phase is extracted with 10 ml. of ethyl acetate, extracted with ethyl acetate and allowed to stand overnight. Precipitation with petroleum ether gives L-2-D-1-amino-1-ethoxycarbonyl-1-methylvinylamino-2-p-ethoxycarbonylphenylacetamide (1888; b.sub.f = 141 silica gel diethyl-ether=10:10 v/v).

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11- Entry 4 of 11

Index: 0001

May 14, 1978

DOCUMENT-IDENTIFIER: US 0713481 A

TITLE: Acylation with a side penicillanic acid equivalent ring

ABSTRACT:

A method is provided for synthesizing β -lactam antibiotics by enzymatic acylation of the parent β -lactam with an activated derivative of the side chain acid wherein a modulator, which consists of one or more compounds different from the reactants and the reaction product and which suppresses the hydrolysis of the activated derivative of the side chain acid and the desired product more than it suppresses the synthesis of the desired product, is added to the reaction mixture, at the beginning of the reaction process, in a concentration from about 0.2 to 100 times 10^{-3} m.u.m.

BSPB:

The present invention relates to an improved method for enzymatic acylation. In particular, the invention relates to the preparation of β -lactam antibiotics by enzymatic acylation of the parent amino β -lactam moiety with an acylating agent which is an activated derivative of the side chain acid.

BSPB:

Enzymatic production of semisynthetic β -lactam antibiotics by acylation of the parent amino β -lactam moiety with the side chain acid or an activated derivative, such as an amide or an ester thereof, is known e.g. from West German patent application having publication No. 1,103,791, Austrian Patent No. 143,840, Dutch patent application No. 72-24139, West German patent application having publication No. 1,611,615, European patent application having publication No. 839,781, international patent application having publication No. WO 83/01061 and from international patent application having publication No. WO 83/02251.

BSPB:

The parent amino β -lactams such as D-aminopenicillanic acid (D-APA) and D-aminodesacetoxycephalosporanic acid (D-ADCA) are commonly produced by fermentation. Because impurities originating from the fermentation, the resulting crude product typically contains unwanted traces of the β -lactam which is used as starting material at a concentration of 10^{-3} to 10^{-4} m.u.m. The crude solution can be purified and crystallized to obtain pure D-APA or D-ADCA. In the D-ADCA case, the fermented penicillin has to go through a rearrangement process before the hydrolysis step.

BSPB:

A drawback of the known methods for enzymatic production of β -lactam antibiotics is that acylation of the parent amino β -lactam with an activated derivative of the side chain acid is that under the reaction conditions used part of the acylating agent hydrolyzes before it has reacted with the amino β -lactam. Thus, when the amide of the side chain acid is used as acylating agent, some free side chain acid and an equivalent amount of ammonia will be generated in the reaction mixture as a result of this hydrolysis. Similarly, when an ester of the side chain acid is used as acylating agent, some free side chain acid and an equivalent amount of the alcohol corresponding to the ester will be generated in the reaction mixture as a

1. *Journal of the American Medical Association*, 1997; 278: 1039-1044.

1997, 1998, 1999, 2000, 2001, 2002, 2003, 2004, 2005, 2006, 2007, 2008, 2009, 2010, 2011, 2012, 2013, 2014, 2015, 2016, 2017, 2018, 2019, 2020, 2021, 2022, 2023, 2024, 2025, 2026, 2027, 2028, 2029, 2030, 2031, 2032, 2033, 2034, 2035, 2036, 2037, 2038, 2039, 2040, 2041, 2042, 2043, 2044, 2045, 2046, 2047, 2048, 2049, 2050, 2051, 2052, 2053, 2054, 2055, 2056, 2057, 2058, 2059, 2060, 2061, 2062, 2063, 2064, 2065, 2066, 2067, 2068, 2069, 2070, 2071, 2072, 2073, 2074, 2075, 2076, 2077, 2078, 2079, 2080, 2081, 2082, 2083, 2084, 2085, 2086, 2087, 2088, 2089, 2090, 2091, 2092, 2093, 2094, 2095, 2096, 2097, 2098, 2099, 2100, 2101, 2102, 2103, 2104, 2105, 2106, 2107, 2108, 2109, 2110, 2111, 2112, 2113, 2114, 2115, 2116, 2117, 2118, 2119, 2120, 2121, 2122, 2123, 2124, 2125, 2126, 2127, 2128, 2129, 2130, 2131, 2132, 2133, 2134, 2135, 2136, 2137, 2138, 2139, 2140, 2141, 2142, 2143, 2144, 2145, 2146, 2147, 2148, 2149, 2150, 2151, 2152, 2153, 2154, 2155, 2156, 2157, 2158, 2159, 2160, 2161, 2162, 2163, 2164, 2165, 2166, 2167, 2168, 2169, 2170, 2171, 2172, 2173, 2174, 2175, 2176, 2177, 2178, 2179, 2180, 2181, 2182, 2183, 2184, 2185, 2186, 2187, 2188, 2189, 2190, 2191, 2192, 2193, 2194, 2195, 2196, 2197, 2198, 2199, 2200, 2201, 2202, 2203, 2204, 2205, 2206, 2207, 2208, 2209, 2210, 2211, 2212, 2213, 2214, 2215, 2216, 2217, 2218, 2219, 2220, 2221, 2222, 2223, 2224, 2225, 2226, 2227, 2228, 2229, 2230, 2231, 2232, 2233, 2234, 2235, 2236, 2237, 2238, 2239, 2240, 2241, 2242, 2243, 2244, 2245, 2246, 2247, 2248, 2249, 2250, 2251, 2252, 2253, 2254, 2255, 2256, 2257, 2258, 2259, 2260, 2261, 2262, 2263, 2264, 2265, 2266, 2267, 2268, 2269, 2270, 2271, 2272, 2273, 2274, 2275, 2276, 2277, 2278, 2279, 2280, 2281, 2282, 2283, 2284, 2285, 2286, 2287, 2288, 2289, 2290, 2291, 2292, 2293, 2294, 2295, 2296, 2297, 2298, 2299, 2300, 2301, 2302, 2303, 2304, 2305, 2306, 2307, 2308, 2309, 2310, 2311, 2312, 2313, 2314, 2315, 2316, 2317, 2318, 2319, 2320, 2321, 2322, 2323, 2324, 2325, 2326, 2327, 2328, 2329, 2330, 2331, 2332, 2333, 2334, 2335, 2336, 2337, 2338, 2339, 2340, 2341, 2342, 2343, 2344, 2345, 2346, 2347, 2348, 2349, 2350, 2351, 2352, 2353, 2354, 2355, 2356, 2357, 2358, 2359, 2360, 2361, 2362, 2363, 2364, 2365, 2366, 2367, 2368, 2369, 2370, 2371, 2372, 2373, 2374, 2375, 2376, 2377, 2378, 2379, 2380, 2381, 2382, 2383, 2384, 2385, 2386, 2387, 2388, 2389, 2390, 2391, 2392, 2393, 2394, 2395, 2396, 2397, 2398, 2399, 2400, 2401, 2402, 2403, 2404, 2405, 2406, 2407, 2408, 2409, 2410, 2411, 2412, 2413, 2414, 2415, 2416, 2417, 2418, 2419, 2420, 2421, 2422, 2423, 2424, 2425, 2426, 2427, 2428, 2429, 2430, 2431, 2432, 2433, 2434, 2435, 2436, 2437, 2438, 2439, 2440, 2441, 2442, 2443, 2444, 2445, 2446, 2447, 2448, 2449, 2450, 2451, 2452, 2453, 2454, 2455, 2456, 2457, 2458, 2459, 2460, 2461, 2462, 2463, 2464, 2465, 2466, 2467, 2468, 2469, 2470, 2471, 2472, 2473, 2474, 2475, 2476, 2477, 2478, 2479, 2480, 2481, 2482, 2483, 2484, 2485, 2486, 2487, 2488, 2489, 2490, 2491, 2492, 2493, 2494, 2495, 2496, 2497, 2498, 2499, 2500, 2501, 2502, 2503, 2504, 2505, 2506, 2507, 2508, 2509, 2510, 2511, 2512, 2513, 2514, 2515, 2516, 2517, 2518, 2519, 2520, 2521, 2522, 2523, 2524, 2525, 2526, 2527, 2528, 2529, 2530, 2531, 2532, 2533, 2534, 2535, 2536, 2537, 2538, 2539, 2540, 2541, 2542, 2543, 2544, 2545, 2546, 2547, 2548, 2549, 2550, 2551, 2552, 2553, 2554, 2555, 2556, 2557, 2558, 2559, 2560, 2561, 2562, 2563, 2564, 2565, 2566, 2567, 2568, 2569, 2570, 2571, 2572, 2573, 2574, 2575, 2576, 2577, 2578, 2579, 2580, 2581, 2582, 2583, 2584, 2585, 2586, 2587, 2588, 2589, 2590, 2591, 2592, 2593, 2594, 2595, 2596, 2597, 2598, 2599, 2600, 2601, 2602, 2603, 2604, 2605, 2606, 2607, 2608, 2609, 2610, 2611, 2612, 2613, 2614, 2615, 2616, 2617, 2618, 2619, 2620, 2621, 2622, 2623, 2624, 2625, 2626, 2627, 2628, 2629, 2630, 2631, 2632, 2633, 2634, 2635, 2636, 2637, 2638, 2639, 2640, 2641, 2642, 2643, 2644, 2645, 2646, 2647, 2648, 2649, 2650, 2651, 2652, 2653, 2654, 2655, 2656, 2657, 2658, 2659, 2660, 2661, 2662, 2663, 2664, 2665, 2666, 2667, 2668, 2669, 2670, 2671, 2672, 2673, 2674, 2675, 2676, 2677, 2678, 26

1. $\frac{1}{2}$ of the total weight of the sample is made up of water.
 2. The weight of the sample is 100 g.
 3. The weight of the sample is 100 g.

[illegible][illegible]

Figure 1

[illegible]

Further new amino .beta.-lactam patent applications having priority to U.S. Pat. 3,174,417, May 1966, and others. Another U.S. Pat. 3,174,418, May 1966, and corresponding foreign patent applications having priority to U.S. Pat. 3,174,418. The hydrolytic enzyme is commercially available. The enzyme may also be a so-called amino acid hydrolase, amylase or amidase. In this connection, reference is, inter alia, made to Hacks to Hays 38-1947, 216 et seq., the contents of which is incorporated by reference.

1914:

It is preferred that the enzyme be a so-called amino acid hydrolase, for example, an amino acid hydrolase. In this connection, reference is made to Hacks to Hays 38-1947, 216 et seq., the contents of which is incorporated by reference. The enzyme is commercially available from Boehringer Mannheim GmbH, Germany, under the trade name Bactol.

1915:

The solubility of the acylating agent such as the L-phenylglycine or L-tyrosine or phenylglycine derivative is a function of the solvent as described in 1914-1915, will vary with the identity of the derivative and with the composition of the reaction medium. In an aqueous system as used in the examples, the solubility of the hydrochloride of L-phenylglycine amide is typically approximately 400 mM. However, the solubility is very dependent on the salt components in the solution, as well as on the pH value and the temperature of the solution. In some embodiments of the process of this invention, the initial reaction mixture is a slurry containing undissolved acylating agent and/or amino .beta.-lactam, which will dissolve partly or fully during the course of the reaction. The .beta.-lactam antibiotic formed may precipitate during the reaction and, also, the hydrolysis products of the acylating agent such as L-phenylglycine and L-tyrosine may precipitate. Hence, in some cases the reaction mixture will be a slurry throughout the duration of the reaction.

1916:

The amino .beta.-lactam, for example 6-APA or 7-ADCA, used in the process of this invention may be obtained by enzymatic hydrolysis of the fermented penicillins or cephalosporins (for example penicillin V, penicillin G or cephalosporin C), or their ring-enlarged analogues (for example 7-ACA and 7-ADCA) or by other means such as acid hydrolysis. In some cases, the crude solution can be used directly without further purification or dilution.

1917:

Generally, the reaction temperature of the process of this invention may vary between about 0.degree. C. and about 30.degree. C., especially between about 5.degree. C. and about 25.degree. C. Temperatures in the range about 25.degree. C.-30.degree. C. may be preferred for convenient operation. The pH value which is optimal depends on the type and purity of enzyme. Using amylase as an example, the optimal pH value is typically in the range from about 6.0 to about 7.0, preferably in the range from about 6.1 to about 6.5. In the preparation of amoxicillin, a pH value in the range from about 6.5 to about 6.9 is preferred. Control of the pH value may be used. Suitable reaction times are from several minutes to several hours, in particular from about 1 hour to about 4 hours. Suitable enzyme concentrations may be from about 1 mM to about 100 mM of the unit of enzyme activity, see below.

1918:

Using the method according to this invention, unusually high yields of the amino .beta.-lactam antibiotic can be obtained. The high yields are obtained when the conditions of this invention are properly selected, the concentration of the acylating agent, the ratio between the concentration of acylating agent and the starting amino .beta.-lactam, the pH value, the enzyme and the identity and amount of mediator.

1919:

7-APA is 7-p-hydroxyphenylglycine amide, 7-HPG is 7-p-hydroxyphenylglycine, 6-APA is 6-p-aminopenicillanic acid, Amox is amoxicillin, Phox is phenoxycarboxylic acid.

and, and 10% to penicillin G area.

[EPR]:

The ratio X is defined as the molar ratio of D-HPG consumed per mole of Amox produced. For practical use this can be transformed to X molar D-HPG per mole Amox, wherein " X molar D-HPG" is the molar amount of D-HPG present at any time during reaction, and the desired product and "molar Amox" is the molar amount of Amox present in the reaction mixture. Thus, if X is 1 this means that only the desired synthesis takes place, no hydrolysis. If X is 2, this means that D-HPG and Amox are formed in equal amounts (on a molar basis). If X is 3, this means that twice as much D-HPG as Amox is present in the reaction mixture (on a molar basis). The ratio X can be calculated at any time during reaction, but in the following examples X is calculated at the reaction stop time, which is defined as the time at which 90% of the theoretical yield of Amox is present in the reaction mixture (based on the inserted amount of 4-AFA). Square brackets are used to designate molar concentrations.

[EPR]:

The following definition of penicillin G arylase activity is used: one unit of arylase is the amount of enzyme that hydrolyzes per minute 1.0 mmole of penicillin G under standard conditions: 50 penicillin G, 0.05M sodium phosphate buffer, pH 7.0, 25 degree C.

[EPR]:

The ratio X is defined as the molar ratio of D-HPG consumed per mole of Amox produced. For practical use this can be transformed to X molar D-HPG per mole Amox, wherein " X molar D-HPG" is the molar amount of D-HPG present at any time during reaction, and the desired product and "molar Amox" is the molar amount of Amox present in the reaction mixture. Thus, if X is 1 this means that only the desired synthesis takes place, no hydrolysis. If X is 2, this means that D-HPG and Amox are formed in equal amounts (on a molar basis). If X is 3, this means that twice as much D-HPG as Amox is present in the reaction mixture (on a molar basis). The ratio X can be calculated at any time during reaction, but in the following examples X is calculated at the reaction stop time, which is defined as the time at which 90% of the theoretical yield of Amox is present in the reaction mixture (based on the inserted amount of 4-AFA). Square brackets are used to designate molar concentrations.

[EPR]:

Retention times in minutes: 2.6 (D-HPG), 3.5 (D-HPGA), 5.0 (6-AFA), 13.5 (Amox).

[EPR]:

In Examples 1-4 the following standard conditions for enzymatic amoxicillin synthesis have been used (see patent application No. WO 92/01161 for further details):

[EPR]:

During the reactions, the pH value of the reaction mixtures was kept constant by titration with 0.1M sodium hydroxide.

[EPR]:

A standard synthesis (immobilized penicillin G arylase from *E. coli*; enzyme using 5.0 U/ml) was carried out with no Phox added. The Phox level was 0.0 mmM in the reaction mixture due to a residual Phox content of 0.0008 mmM in the enzyme. The results are reported in Table 1.

[EPR]:

A standard synthesis (immobilized penicillin G arylase from *E. coli*; enzyme using 5.0 U/ml) was carried out with no Phox added. The Phox level in the reaction mixture was 0.0 mmM. The results are reported in Table 1.

[EPR]:

Four different immobilized pen G arylase preparations were used: a) immobilized penicillin G arylase from *E. coli*; enzyme using 5.0 U/ml; b) immobilized penicillin G arylase from *E. coli*; enzyme using 1.0 U/ml; c) immobilized penicillin G arylase from *E. coli*; enzyme using 0.1 U/ml; and d) immobilized penicillin G arylase from *E. coli*; enzyme using 0.01 U/ml.

[EPR]:

Various pen G arylase preparations were obtained from *E. coli* (Gen. No. 100000, ca. 100 U/ml, and from *EPH*, ca. 10 U/ml. An enzyme dosage of 5.0 U/ml were applied in the two series. The results are reported in Table 1.

TABLE 3:

Enzymatic Synthesis of Amoxicillin Using Immobilized Penicillin G Acylase from *E. coli*; enzyme dosage: 0.1 mg/ml. The reaction was carried out with an L- -threonine acid 100-1. The Phox concentration level was 0.1 mM in the reaction mixture. The results obtained are reported in Table 3.

TABLE 4:

Enzymatic Synthesis of Amoxicillin Using Immobilized Penicillin G Acylase, enzyme dosage: 0.1 mg/ml. The reaction was carried out with an L- -threonine acid 100-1. The Phox concentration level in the reaction mixture was 0.1 mM. The results obtained are reported in Table 4.

TABLE 5:

The data were obtained to improve synthesis performance. Standard synthesis conditions, and immobilized Penicillin G Acylase from *E. coli*; enzyme dosage: 0.1 mg/ml. In the cases that when they are present in the reaction mixture in a concentration within the interval specified, synthesis of the desired product is maintained as compared to hydrolysis of the acylating agent:

DEFC:

Enzyme Activity

DEFC:

Enzymatic Synthesis of Amoxicillin Using a Fixed Dosage of Immobilized Pen G Acylase and Varying the Phox Concentration in the Reaction Mixture from 2.6 to 0.1 mM

DEFC:

Enzymatic Synthesis of Amoxicillin Using a Fixed Dosage of Immobilized Pen G Acylase and Varying the Phyl Concentration in the Reaction Mixture from 32.9 to 0.1 mM

DEFC:

Enzymatic Synthesis of Amoxicillin Using Various Immobilized Preparations of Pen G Acylase and Varying the Phox Concentration in the Reaction Mixture from 0.1 to 32.9 mM

DEFC:

Enzymatic Synthesis of Amoxicillin Using Varying Amounts of Enzyme and a Constant Concentration of Phox

DEFC:

Enzymatic Synthesis of Amoxicillin by Using Soluble Pen G Acylase (Two Different Suppliers) and Varying the Phox Concentration in the Reaction Mixture from 0.1 to 32.9 mM

DEFC:

Enzymatic Synthesis of Amoxicillin by Using Immobilized Pen G Acylase and Varying the Concentration of L- -Threonine Acid from 0.1 to 100-1 mM

DEFC:

Enzymatic Synthesis of Amoxicillin by Using Immobilized Pen G Acylase and Varying the 2-Thiophenecarboxylic Acid Concentration from 0.1 to 0.3 mM in the Reaction Mixture

DEFC:

Enzymatic Synthesis of Amoxicillin Using Immobilized Pen G Acylase and Varying the Enzyme Dosage

DEFC:

TABLE 4
Enzymatic Synthesis of Amoxicillin Using Immobilized Pen G Acylase and Varying the 2-Thiophenecarboxylic Acid Concentration from 0.1 to 0.3 mM in the Reaction Mixture
Experiment mixture 1 ml Amox 2-HPS time

----- A 31 11.3 1.17 0.01 1.75 B 31 23.5 1.20

1. A method for providing a semisynthetic beta-lactam antibiotic by enzyme

TABLE 1
Initial velocity Enzyme (U/mg)
Time (min) 0 10 20 30 40 50 60 70 80 90 100
Reaction Mixture (ml) 1.0 1.0 1.0 1.0 1.0 1.0 1.0 1.0 1.0 1.0 1.0
Substrate (mg/ml) 0.1 0.2 0.3 0.4 0.5 0.6 0.7 0.8 0.9 1.0 1.1
Product (mg/ml) 0.0 0.1 0.2 0.3 0.4 0.5 0.6 0.7 0.8 0.9 1.0
Enzyme (mg/ml) 0.1 0.1 0.1 0.1 0.1 0.1 0.1 0.1 0.1 0.1 0.1

2. A method for providing a semisynthetic beta-lactam antibiotic by enzyme
catalyzed acylation of the parent beta-lactam with an amide or ester of the
side chain acid wherein a modulator, which is a carboxylic acid of 2 to 21
carbon atoms, and is different from the reactants and the reaction product is
added to the reaction mixture, at the beginning of the reaction process, in a
concentration of 0.1 to 1.0 mg/ml of the reaction mixture.

3. A method for providing a semisynthetic beta-lactam antibiotic by enzyme
catalyzed acylation of the parent beta-lactam with an amide or ester of the
side chain acid wherein a modulator, which is a carboxylic acid of 2 to 21
carbon atoms, and is different from the reactants and the reaction product is
added to the reaction mixture, at the beginning of the reaction process, in a
concentration of 0.1 to 1.0 mg/ml of the reaction mixture.

4. A method for providing a semisynthetic beta-lactam antibiotic by enzyme
catalyzed acylation of the parent beta-lactam with an amide or ester of the
side chain acid wherein a modulator, which is a carboxylic acid of 2 to 21
carbon atoms, and is different from the reactants and the reaction product is
added to the reaction mixture, at the beginning of the reaction process, in a
concentration of 0.1 to 1.0 mg/ml of the reaction mixture.

WEST

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ACKNOWLEDGMENTS

1. 凡在本行开立存款账户的存款人，均可向本行申请开立支票。
 2. 支票的出票人必须是在本行开立存款账户的存款人。
 3. 支票的金额必须与存款账户的余额相符。
 4. 支票的有效期为自签发之日起10日内。
 5. 支票的收款人必须为本行开户的存款人。
 6. 支票的用途必须符合国家有关规定。
 7. 支票的签发必须使用本行规定的支票格式。
 8. 支票的签发必须加盖本行规定的印章。
 9. 支票的签发必须填写完整。
 10. 支票的签发必须真实。

WEST

Generate Collection

Title: Recovery of Ampicillin

Patent: 4,041,111

Date: 12/1/1973

1. SUMMARY-DESCRIPTION: This is a process for the recovery of ampicillin.

ABSTRACT:

Process for the recovery of ampicillin from a mixture containing ampicillin and D-aminopenicillanic acid (D-APA), in which a mixture of ampicillin and D-APA, with a pH higher than 7, which apart from any solid ampicillin being present is homogeneous at a pH between 7 and 8.5, is subjected to a pH lowering till a pH lower than 6.2 is reached, and the solid substance present is recovered. The process is in particular suitable to be applied to the recovery of a mixture which is obtained after the enzymatic acylation reaction of D-APA with a penicillanic acid derivative as acylation agent. Pure ampicillin can thus be recovered in a simple way.

BACKGROUND:

The invention relates to a process for the recovery of ampicillin from a mixture containing ampicillin and 6-aminopenicillanic acid (6-APA).

OBJECTS:

In the preparation of ampicillin, with D-APA being acylated with a penicillanic acid derivative, the recovery of the ampicillin and working up of the reaction mixture are difficult in general.

DISCUSSION:

A process for isolating the ampicillin pure from a mixture containing ampicillin and minor quantities of D-APA is described in JP-A-47030667. According to the process described in this Japanese publication, an acid aqueous mixture containing D-APA and ampicillin is subjected to an extraction with butanol or isomylalcohol, after which the pH is raised to a value between 6 and 7 and the product is recovered by complete boiling down and freeze-drying. The drawback of this method is that organic solvents that are alien to the process have to be added. In addition, complete boiling down and freeze-drying is not industrially practicable. Moreover, the process involves formation of salts that are included in the freeze-dried product.

DISCUSSION:

JP-A-474411 discloses a process wherein ampicillin is recovered from a mixture of ampicillin and amino-penicillanic acid by conversion of the ampicillin to the trialkylamine salt and recover the ampicillin as its trialkylamine salt.

DISCUSSION:

In view of the above, it is possible to provide a simple, industrially practicable process for the recovery of ampicillin from a mixture with it making use of solid organic solvents that are alien to the process.

DISCUSSION:

This is achieved according to the invention in that a mixture containing ampicillin and D-APA and having a pH higher than 7, which, apart from any solid ampicillin that is present, is homogeneous at a pH between 7 and 8.5, is subjected to a pH lowering to a pH lower than 6.2, and that the solid substance present is recovered.

DISCUSSION:

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1. $\frac{1}{2}$ 2. $\frac{1}{3}$ 3. $\frac{1}{4}$ 4. $\frac{1}{5}$ 5. $\frac{1}{6}$ 6. $\frac{1}{7}$ 7. $\frac{1}{8}$ 8. $\frac{1}{9}$ 9. $\frac{1}{10}$ 10. $\frac{1}{11}$ 11. $\frac{1}{12}$ 12. $\frac{1}{13}$ 13. $\frac{1}{14}$ 14. $\frac{1}{15}$ 15. $\frac{1}{16}$ 16. $\frac{1}{17}$ 17. $\frac{1}{18}$ 18. $\frac{1}{19}$ 19. $\frac{1}{20}$ 20. $\frac{1}{21}$ 21. $\frac{1}{22}$ 22. $\frac{1}{23}$ 23. $\frac{1}{24}$ 24. $\frac{1}{25}$ 25. $\frac{1}{26}$ 26. $\frac{1}{27}$ 27. $\frac{1}{28}$ 28. $\frac{1}{29}$ 29. $\frac{1}{30}$ 30. $\frac{1}{31}$ 31. $\frac{1}{32}$ 32. $\frac{1}{33}$ 33. $\frac{1}{34}$ 34. $\frac{1}{35}$ 35. $\frac{1}{36}$ 36. $\frac{1}{37}$ 37. $\frac{1}{38}$ 38. $\frac{1}{39}$ 39. $\frac{1}{40}$ 40. $\frac{1}{41}$ 41. $\frac{1}{42}$ 42. $\frac{1}{43}$ 43. $\frac{1}{44}$ 44. $\frac{1}{45}$ 45. $\frac{1}{46}$ 46. $\frac{1}{47}$ 47. $\frac{1}{48}$ 48. $\frac{1}{49}$ 49. $\frac{1}{50}$ 50. $\frac{1}{51}$ 51. $\frac{1}{52}$ 52. $\frac{1}{53}$ 53. $\frac{1}{54}$ 54. $\frac{1}{55}$ 55. $\frac{1}{56}$ 56. $\frac{1}{57}$ 57. $\frac{1}{58}$ 58. $\frac{1}{59}$ 59. $\frac{1}{60}$ 60. $\frac{1}{61}$ 61. $\frac{1}{62}$ 62. $\frac{1}{63}$ 63. $\frac{1}{64}$ 64. $\frac{1}{65}$ 65. $\frac{1}{66}$ 66. $\frac{1}{67}$ 67. $\frac{1}{68}$ 68. $\frac{1}{69}$ 69. $\frac{1}{70}$ 70. $\frac{1}{71}$ 71. $\frac{1}{72}$ 72. $\frac{1}{73}$ 73. $\frac{1}{74}$ 74. $\frac{1}{75}$ 75. $\frac{1}{76}$ 76. $\frac{1}{77}$ 77. $\frac{1}{78}$ 78. $\frac{1}{79}$ 79. $\frac{1}{80}$ 80. $\frac{1}{81}$ 81. $\frac{1}{82}$ 82. $\frac{1}{83}$ 83. $\frac{1}{84}$ 84. $\frac{1}{85}$ 85. $\frac{1}{86}$ 86. $\frac{1}{87}$ 87. $\frac{1}{88}$ 88. $\frac{1}{89}$ 89. $\frac{1}{90}$ 90. $\frac{1}{91}$ 91. $\frac{1}{92}$ 92. $\frac{1}{93}$ 93. $\frac{1}{94}$ 94. $\frac{1}{95}$ 95. $\frac{1}{96}$ 96. $\frac{1}{97}$ 97. $\frac{1}{98}$ 98. $\frac{1}{99}$ 99. $\frac{1}{100}$ 100. $\frac{1}{101}$ 101. $\frac{1}{102}$ 102. $\frac{1}{103}$ 103. $\frac{1}{104}$ 104. $\frac{1}{105}$ 105. $\frac{1}{106}$ 106. $\frac{1}{107}$ 107. $\frac{1}{108}$ 108. $\frac{1}{109}$ 109. $\frac{1}{110}$ 110. $\frac{1}{111}$ 111. $\frac{1}{112}$ 112. $\frac{1}{113}$ 113. $\frac{1}{114}$ 114. $\frac{1}{115}$ 115. $\frac{1}{116}$ 116. $\frac{1}{117}$ 117. $\frac{1}{118}$ 118. $\frac{1}{119}$ 119. $\frac{1}{120}$ 120. $\frac{1}{121}$ 121. $\frac{1}{122}$ 122. $\frac{1}{123}$ 123. $\frac{1}{124}$ 124. $\frac{1}{125}$ 125. $\frac{1}{126}$ 126. $\frac{1}{127}$ 127. $\frac{1}{128}$ 128. $\frac{1}{129}$ 129. $\frac{1}{130}$ 130. $\frac{1}{131}$ 131. $\frac{1}{132}$ 132. $\frac{1}{133}$ 133. $\frac{1}{134}$ 134. $\frac{1}{135}$ 135. $\frac{1}{136}$ 136. $\frac{1}{137}$ 137. $\frac{1}{138}$ 138. $\frac{1}{139}$ 139. $\frac{1}{140}$ 140. $\frac{1}{141}$ 141. $\frac{1}{142}$ 142. $\frac{1}{143}$ 143. $\frac{1}{144}$ 144. $\frac{1}{145}$ 145. $\frac{1}{146}$ 146. $\frac{1}{147}$ 147. $\frac{1}{148}$ 148. $\frac{1}{149}$ 149. $\frac{1}{150}$ 150. $\frac{1}{151}$ 151. $\frac{1}{152}$ 152. $\frac{1}{153}$ 153. $\frac{1}{154}$ 154. $\frac{1}{155}$ 155. $\frac{1}{156}$ 156. $\frac{1}{157}$ 157. $\frac{1}{158}$ 158. $\frac{1}{159}$ 159. $\frac{1}{160}$ 160. $\frac{1}{161}$ 161. $\frac{1}{162}$ 162. $\frac{1}{163}$ 163. $\frac{1}{164}$ 164. $\frac{1}{165}$ 165. $\frac{1}{166}$ 166. $\frac{1}{167}$ 167. $\frac{1}{168}$ 168. $\frac{1}{169}$ 169. $\frac{1}{170}$ 170. $\frac{1}{171}$ 171. $\frac{1}{172}$ 172. $\frac{1}{173}$ 173. $\frac{1}{174}$ 174. $\frac{1}{175}$ 175. $\frac{1}{176}$ 176. $\frac{1}{177}$ 177. $\frac{1}{178}$ 178. $\frac{1}{179}$ 179. $\frac{1}{180}$ 180. $\frac{1}{181}$ 181. $\frac{1}{182}$ 182. $\frac{1}{183}$ 183. $\frac{1}{184}$ 184. $\frac{1}{185}$ 185. $\frac{1}{186}$ 186. $\frac{1}{187}$ 187. $\frac{1}{188}$ 188. $\frac{1}{189}$ 189. $\frac{1}{190}$ 190. $\frac{1}{191}$ 191. $\frac{1}{192}$ 192. $\frac{1}{193}$ 193. $\frac{1}{194}$ 194. $\frac{1}{195}$ 195. $\frac{1}{196}$ 196. $\frac{1}{197}$ 197. $\frac{1}{198}$ 198. $\frac{1}{199}$ 199. $\frac{1}{200}$ 200. $\frac{1}{201}$ 201. $\frac{1}{202}$ 202. $\frac{1}{203}$ 203. $\frac{1}{204}$ 204. $\frac{1}{205}$ 205. $\frac{1}{206}$ 206. $\frac{1}{207}$ 207. $\frac{1}{208}$ 208. $\frac{1}{209}$ 209. $\frac{1}{210}$ 210. $\frac{1}{211}$ 211. $\frac{1}{212}$ 212. $\frac{1}{213}$ 213. $\frac{1}{214}$ 214. $\frac{1}{215}$ 215. $\frac{1}{216}$ 216. $\frac{1}{217}$ 217. $\frac{1}{218}$ 218. $\frac{1}{219}$ 219. $\frac{1}{220}$ 220. $\frac{1}{221}$ 221. $\frac{1}{222}$ 222. $\frac{1}{223}$ 223. $\frac{1}{224}$ 224. $\frac{1}{225}$ 225. $\frac{1}{226}$ 226. $\frac{1}{227}$ 227. $\frac{1}{228}$ 228. $\frac{1}{229}$ 229. $\frac{1}{230}$ 230. $\frac{1}{231}$ 231. $\frac{1}{232}$ 232. $\frac{1}{233}$ 233. $\frac{1}{234}$ 234. $\frac{1}{235}$ 235. $\frac{1}{236}$ 236. $\frac{1}{237}$ 237. $\frac{1}{238}$ 238. $\frac{1}{239}$ 239. $\frac{1}{240}$ 240

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|------|------|------|------|------|
| 1990 | 1991 | 1992 | 1993 | 1994 |
| 1.0 | 1.0 | 1.0 | 1.0 | 1.0 |
| 1.0 | 1.0 | 1.0 | 1.0 | 1.0 |
| 1.0 | 1.0 | 1.0 | 1.0 | 1.0 |

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1. *Journal of the American Medical Association*, 1997; 278: 1039-1044.

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$\frac{1}{2} \times \frac{1}{2} = \frac{1}{4}$

[illegible]

1. A process for recovering penicillin from

claim 1,

comprising the steps of:

1.1. Enzymatic coupling of 2.0 mM of PGA and 2.0 mM of 6-APA at 5.degree. C.,

followed by working up.

1.2. After 1 hour the pH had risen to 6.0. By means of concentrated aqueous

NH.sub.4 the pH was brought to 5.6 and after 5 minutes the reaction mixture was filtered through a G-3 glass filter; the residue was washed with 100 ml of water (5.degree. C.). This residue was a mixture of enzyme and PG formed during the reaction.

1.3. After 1 hour the pH had risen to 6.0. By means of concentrated aqueous

NH.sub.4 the pH was brought to 5.6 and after 5 minutes the reaction mixture was filtered through a G-3 glass filter; the residue was washed with 100 ml of water (5.degree. C.). This residue was a mixture of enzyme and PG formed during the reaction.

1.4. Enzymatic coupling of 2.0 mM of PGA and 2.0 mM of 6-APA at 5.degree. C.,

followed by working up.

2. A process for recovering penicillin from a mixture containing ampicillin

and 6-aminopenicillanic acid (6-APA) comprising the steps of:

2.1. A process according to claim 1, wherein the mixture contains 1-50 mol % of

6-APA, calculated relative to the total amount of 6-APA and ampicillin.

2.2. A process according to claim 1, wherein the mixture contains 1-50 mol % of

6-APA, calculated relative to the total amount of 6-APA and ampicillin.

2.3. A process according to claim 1, wherein the mixture contains 1-50 mol % of

6-APA, calculated relative to the total amount of 6-APA and ampicillin.

2.4. A process according to claim 1, wherein the mixture contains 1-50 mol % of

6-APA, calculated relative to the total amount of 6-APA and ampicillin.

2.5. A process according to claim 1, wherein the mixture contains 1-50 mol % of

6-APA, calculated relative to the total amount of 6-APA and ampicillin.

2.6. A process according to claim 1, wherein the mixture contains 1-50 mol % of

6-APA, calculated relative to the total amount of 6-APA and ampicillin.

2.7. A process according to claim 1, wherein the mixture contains 1-50 mol % of

6-APA, calculated relative to the total amount of 6-APA and ampicillin.

STEP:

1. Aqueous suspension of ampicillin, wherein the ampicillin is acylated, is added to the pH of the mixture by 20% sodium hydroxide and water, and the pH is adjusted to a pH of approximately 6.0-7.0 and then.

STEP:

providing the mixture containing ampicillin and D-APA, said mixture having an initial pH greater than 7 and reducing the pH to a pH between 7 and 8.5 and then adjusting the pH to approximately 6.0-7.0 and then.

STEP:

1. When the initial pH of the mixture is a pH value above 8.5 and then reducing the pH to approximately 6.0-7.0 and then.

STEP:

obtaining a mixture containing ampicillin and D-aminopenicillin that is prepared from the reaction mixture of an enzymatic acylation reaction in which D-APA is acylated using D-phenylglycineamide (PGA) or esters of D-phenylglycine, said mixture having an initial pH greater than 7; and

STEP:

reducing the pH of said mixture and crystallizing out ampicillin.

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1. 在 1990 年 1 月 1 日以前，《中华人民共和国民法通则》 第 134 条第 2 款规定：“有民事行为能力的人，有法律规定的民事权利能力和民事行为能力，依法独立享有民事权利和承担民事义务。”

This invention relates to a process for the preparation of β -lactam derivatives by enzymatic acylation of the parent amino β -lactam with an acylating agent. The amino β -lactam may be 6-aminopenicillanic acid (6-APA), 6-aminodesacetoxycephalosporanic acid (7-ADCA), 7-aminocephalexanic acid (7-ACA) or 7-amino-6-chloro-3-cephem-4-carboxylate and the acylating agent may be a derivative of D-phenylglycine or D-phenylalanine.

Today, semisynthetic β -lactam antibiotics such as Ampicillin, Amoxicillin, Cefadex, Cephalexin, Cephadroxil and Cephadrilysin are prepared in industry by chemical methods, for example by reacting an amino β -lactam such as D-phenylglycyl-L-proline, usually having its carboxyl group protected, with an activated side chain derivative, followed by the removal of the protecting group by hydrolysis. It is important due to, for example, yield, that the amino β -lactam, for example D-APA, is used in a pure, dry form, preferably in a purity higher than 95%. For example, Ampicillin (D-2-amino-6-aminophenylglycyl-D-phenylglycyl-L-proline) can be prepared by reacting D-APA, having a carboxyl group protected, with D-phenylglycyl-L-proline, followed by removal of the protecting group by hydrolysis. These reactions typically involve costly steps such as sublimation before the reaction and various solvents like methylene chloride and dimethyl sulfoxide.

Hydroxylation of Anipiline from pure p-AFA and 1-Hydroxyglycine derivatives such as a lower alkyl ester is known from West German patent application having publication No. 1,100,000, Austrian Patent No. 349,484, French patent application No. 1,100,000, West German patent application having publication No. 1,100,000 and European patent application having publication No. 1,100,000, however, described in the prior art have typically used a low amount of the 1-Hydroxyglycine derivative and below 4 mM of p-AFA, the highest yield reported was 44%. European patent application having publication No. 1,100,000.

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1.1.1. preferred for the enzyme in a suitable form, for example, as a suspension in water or in an immiscible solvent. Any of the various immobilization techniques for enzyme is commercially available from Boehringer Mannheim GmbH, Germany, under the trade name Biopack.

DEPR:

The solubility of the acylating agent such as the L-phenylglycine or L-tyrosine or phenylglycine derivative will vary with the identity of the enzyme and with the composition of the reaction medium. In an aspect of the invention, for example, the solubility of the enzyme in a medium is about 10 mg/ml at pH 7. However, the solubility is very dependent on the salt concentration in the solution, as well as on the pH value and the temperature of the solution. In some embodiments of the process of this invention, the initial reaction mixture is a slurry containing undissolved acylating agent and/or β -lactam, which will dissolve partly or fully during the course of the reaction. The β -lactam formed may precipitate during the reaction and, also, the hydrolysis products of the acylating agent such as L-phenylglycine and L-tyrosine or phenylglycine may precipitate. Hence, in many cases the reaction mixture will be a slurry throughout the reaction.

DEPR:

The amino β -lactam, for example D-ALA or V-ACA, used in the process of this invention may be obtained by enzymatic hydrolysis of the fermented penicillins or cephalosporins, (for example penicillin V, penicillin G or cephalosporin C) or their ring enlarged analogues (for example V-ACA and G-ACA) or derivatives thereof followed by removal of the hydrolysis by-product, if desired (phenoxycetic acid etc.). Advantageously, the crude solution can be used directly without further purification or dilution.

DEPR:

Generally, the reaction temperature of the process of this invention may vary between about 0.degree. C. and about 35.degree. C., is especially between about 15.degree. C. and about 30.degree. C. Temperatures in the range about 15.degree. C.-30.degree. C. may be preferred for convenient operation. The suitable pH value depends on the type and purity of enzyme. Using Escherichia coli enzyme, the pH value is typically in the range from about 5.5 through about 7.5, preferably in the range from about 6.1 through about 7. For the preparation of Amoxicillin, a pH value in the range from about 5.5 through about 6.4 is preferred. Control of the pH value may be used. Suitable reaction times are from several minutes to several hours, in particular from about 1 hour to about 4 hours. Suitable enzyme concentrations may be from about 1 mg/ml to about 10 mg/ml of the unit of enzyme activity, see below.

DEPR:

Using the process according to this invention, extraordinary high yields can be obtained. The high yields are obtained using the teachings of this invention and properly selecting the concentration of the acylating agent, the ratio between the concentration of acylating agent and the starting amino β -lactam, the pH value and the enzyme.

DEPR:

An embodiment of penicillin G acylase activity for all units is used: the unit is defined as the amount of enzyme that hydrolyses per minute 1.0 mole penicillin G under standard conditions: penicillin G, 0.1M sodium phosphate buffer, pH value 6.1, 25.degree. C.

DEPR:

A solution of 1.0 mM D-ALA and 1.0 mM V-ACA in a concentration as indicated in table 1 is adjusted to pH value 6.4 and equilibrated at 25.degree. C. and 400 rpm in a shaking bath. Escherichia coli, supplied from Boehringer Mannheim GmbH, Germany, is added, for example, 10 mg/ml of the unit of enzyme activity.

DEPR:

Example 1 is repeated in Example 3, only 1.5 mM γ -ABA is used instead of 1 mM. The reaction mixture is analysed for product and the maximal yields obtained at different temperatures of 1-4 hours at 37°C are shown in Table 3.

IEPB:

150 mM γ -ABA and 7.5 mM D-PGA sulphate salt are adjusted to a pH value as indicated in Table 3, and the enzymatic synthesis is carried out at 37 degree. 1.5 mg pH-stat-machine, total volume 1.5 ml and 750 U soluble enzyme from Escherichia coli.

IEPB:

Starting with 150 mM γ -ABA and 7.5 mM D-PGA at pH value 6.4 and 750 U soluble enzyme from Escherichia coli, total volume: 1.5 ml and running the synthesis at temperatures as indicated in Table 4, the maximal yields of Ampicillin obtained are shown in Table 4.

IEPB:

This example was performed analogously with Example 1 using D-PGA instead of P-PGA. The maximal yields of Ampicillin obtained are as stated in Table 5.

IEPB:

Pen V acetyl penicillin from penicillium is purified by filtration, extraction and acetyl acetate and 10% NaOH aqueous phase resulting in a solution of 1 weight/volume pen V is hydrolysed by Semaclyase TM, immobilised pen V acylase from Novo Nordisk A/S at a pH value of 7.0. The phenoxyacetic acid is removed by extraction and to the resulting 6-APA (150 mM) solution, containing minor amounts of biproducts from degraded pen V and 6-APA, is added 45 U/ml soluble enzyme from Escherichia coli and D-PGA (to a final concentration of 7.5 mM). The pH value is adjusted to 6.4 and the reaction is allowed to proceed at 37 degree, 3, keeping the pH value constant.

IEPB:

Under these conditions a total of 135 mmole of Ampicillin (90 %) is formed per liter of reaction volume.

IEPB:

500 mg of immobilised enzyme is suspended ad 10 ml with water. The enzyme solution was mixed with a solution of γ -ABA and D-PGA to a total volume of 25 ml the reaction mixture containing 135 mM γ -ABA and 7.5 mM D-PGA, having pH value 6.4 and purified at room temperature. The synthesis reaction was allowed to proceed at pH 6.4 until 12 hours after which 45 U of the γ -ABA was converted to Ampicillin.

IEPB:

A mixture of 969 mg γ -ABA and 3718 mg HPGA in water is adjusted to pH 6.2 at 18 degree, 3, and 1650 U soluble penicillin G acylase from E. coli is added to a final volume of 100 ml. The synthesis is allowed to proceed at constant temperature, using 1M sulphuric acid to keep the pH at 6.2. After 12 hours the reaction mixture contained 135 mM Ampicillin, corresponding to a yield of 135 mmole of Ampicillin consumption.

IEPB:

1650 U soluble penicillin G acylase from E. coli is added to a mixture of γ -ABA and HPGA 135 mM and 135 mM final concentration, respectively in water at pH 6.2 and 18 degree, 3. After reacting for 4 hours keeping the temperature and pH constant using 1M sulphuric acid for the titration, 135 mM Ampicillin was produced 90% yield based on HPLC-analysis.

IEPB:

Starting with 135 mM γ -ABA, 135 mM HPGA, 1650 U soluble penicillin G acylase from E. coli, 135 mM Ampicillin was produced after 4 hours, when the reaction was carried out at pH 6.2 and 18 degree, 3.

IEPB:

Same conditions as described in example 3, using 135 mM γ -ABA and 135 mM HPGA.

wherein the amount of the starting agent is 10-fold to 100-fold greater than the amount of the amino .beta.-lactam.

CLPR:

1. A process according to claim 1 or 2, wherein the acylating agent is propionyl chloride or N-(3-hydroxyphenyl)glycine or derivatives thereof.

CLPR:

2. A process according to claim 1 or 2, wherein the amino .beta.-lactam is penicillin G, penicillin V, penicillin K, penicillin M, penicillin X, penicillin G, penicillin V, penicillin K, penicillin M, or penicillin X.

CLPR:

3. A process according to claim 1 or 2, wherein the concentration of the amino .beta.-lactam in the reaction mixture when the enzymatic reaction starts is in the range from about 5% to about 75% mM.

CLPR:

4. A process according to claim 1 or 2, wherein the concentration of acylating agent in the reaction mixture when the enzymatic reaction starts is greater than 5% mM.

CLPR:

5. A process according to claim 10, wherein the concentration of the acylating agent in the reaction mixture when the enzymatic reaction starts is greater than 5% mM.

CLPR:

6. A process according to claim 1 or 2, wherein the amount of the acylating agent in the reaction mixture when the enzymatic reaction starts is above the solubility of the agent in the reaction mixture.

CLPR:

7. A process according to claim 1 or 2, wherein the amount of the acylating agent in the starting reaction mixture is greater than half the amount of said agent which is soluble in the reaction mixture plus the amount of the amino .beta.-lactam in the reaction mixture when the enzymatic reaction starts.

CLPR:

8. A process according to claim 1 or 2, wherein the enzyme used is classified as EC 3.5.1.11.

CLPR:

9. A process according to claim 1 or 2, wherein the enzyme used is an alkaline hydrolytic penicillinase that is classified as EC 3.5.1.11.

CLPR:

10. A process according to claim 1 or 2, wherein an enzyme in reusable form is used.

CLPR:

11. A process according to claim 1 or 2, wherein the enzymatic reaction is carried out in a batch or continuous system.